

POTENTIAL DRUG TARGETS FOR ALOIN AND MICRODONTIN: AN *IN-SILICO* ANALYSIS

LOKESH RAVI\*, ADHITHYA RAGUNATHAN

Division of Biomolecules and Genetics, School of Biosciences and Technology, VIT University, Vellore, Tamil Nadu, India.  
Email: lokesh.ravi@vit.ac.in

Received: 17 December 2015, Revised and Accepted: 04 March 2016

## ABSTRACT

**Objective:** The aim of this study was to study the interactions of Aloin and microdонтin with antimalarial drug targets.

**Methods:** The ADMET properties of Aloin and microdонтin were analyzed using LigandScout and Osiris Molecular Property Predictor tools. The protein-ligand docking was performed in AutoDock software. AutoDock result was analyzed using PyMol and LigPlot+ software.

**Results:** ADMET analysis suggests no major side effects for both Aloin and microdонтin. Docking results show that Aloin had the highest significance with *Plasmodium falciparum* calcium-dependent protein kinase (PfCDPK2) with a free binding energy of -8.01 Kcal/Mol, Ki value of 1.35  $\mu$ M and 6 hydrogen bonds. Microdонтin had the highest significance toward Glutaredoxin-1 with -8.04 Kcal/Mol, Ki value of 1.28  $\mu$ M and 3 hydrogen bonds.

**Conclusion:** Based on the observed results for the studied drug targets, the proposed mechanism of action of Aloin is suggestively concluded as PfCDPK2 and for microdонтin as glutaredoxin-1.

**Keywords:** Anti-malaria, Aloin, Microdонтin, Autodock, Ligandscout.

## INTRODUCTION

*Aloe vera* predominantly distributed in the tropical and subtropical regions throughout the world is a succulent xerophyte belonging to the family Liliaceae. Though *A. vera* plants are indigenous to Africa now, their existence could be found all over the world [1]. This genus *Aloe* contains more than 600 species [2]. Their water storage tissue is large, and so they are capable of adapting to areas with low water availability [1]. *A. vera* consists of many biologically important compounds. *Aloe percrassa* is, traditionally, used in Ethiopia to treat Malaria; it is also used to treat wounds and gastrointestinal problems; it is locally known as Ere-Senay in Ethiopia. Recently, Gereziher *et al.* evaluated the antimalarial potency of anthrone derivatives and leaf latex extracted from *Aloe percrassa*. Chloroquine-resistant *Plasmodium berghei* was used to infect the test mice. It was reported that anti-malarial activity of Aloin and microdонтin were lower than that of the leaf latex and concluded that the components of leaf latex had synergistic activity [2]. Several anti-malarial studies are being carried out all over the globe using *Aloe* and other medicinal, to compensate the current demand for the effective anti-malarial drug. The mortality rates of malaria have decreased by 47% in the global level and by 54% in the WHO African region; yet, 5,84,000 people died due to malaria in 2013 out of which 78% were children under the age of 5 [3].

Here, in this study, Aloin and microdонтin were docked with some of the known *Plasmodium* proteins to find the potential drug targets using AutoDock. The studied proteins were; *P. falciparum* lactate dehydrogenase (PfLDH) with PDB ID: 2  $\times$  8L. It plays a key catalytic role in the reduction of pyruvate [4]. Plasmeprin-2 with PDB ID: 1LF4. It is an aspartic protease, with an important role in hemoglobin degradation pathway [5]. Thioredoxin-2 with PDB ID: 3UL3, it is a redox protein that guards the plasmodium cells against high fluxes of reactive oxygen species [6]. Glutaredoxin-1 with PDB ID: 4MZC, it is also a redox protein, working against reactive oxygen species [7]. *P. falciparum* protein kinase-5 with PDB ID: 1OB3. Is an important protein kinase in *P. falciparum* and is believed to be essential for regulation of nuclear division [8]. Falcipain-2 with PDB ID: 2GHU. It is a cysteine protease, with key function in degradation of hemoglobin [9]. *P. falciparum* phosphoethanolamine methyltransferase with PDB ID: 3UJ9. It is a

methyltransferase enzyme, with a key function in membrane biogenesis in *Plasmodium*. This protein is unique for *Plasmodium* and hence is an ideal drug target [10]. *P. falciparum* ADP-Ribosylation factor-1 with PDB ID: 3LRP. It is an important vesicular trafficking protein [11]. *P. falciparum* gamete antigen-27 with PDB ID: 1N81. It is a crucial protein involved in the development of the gamete in *P. falciparum* [12].

## METHODS

All protein molecules were downloaded from RCSB Protein Data Bank website. Structure with the following PDB ID were used in this study; 3UL3, 1LF4, 3UJ9, 1OB3, 2X8L, 1N81, 4MVF, 3LRP, 4MZC, 2GHU, 3K7Y. The structure of the Aloin was downloaded from PubChem website, Aloin 313325. The structure of microdонтin was sketched using ChemSketch. Properties of the ligand molecules were analyzed using LigandScout.

Protein-ligand docking was performed using AutoDock. 4.2.1 [13-15], using Generic Algorithm parameters. Binding sites of each protein molecule was predicted using MetaPocket online tool [16,17]. The docking results from AutoDock were saved in pdb file format and were analyzed using PyMol [18,19] and LigPlus software [20,21] to view the docking positions and bonding.

Osiris property explorer an online molecular properties prediction tool was used to analyze the biological properties of Aloin and microdонтin. LigandScout [22,23] was used to analyze the chemical nature and structural arrangement of the ligand molecules.

## RESULTS

LigandScout analysis of properties of Aloin and microdонтin are listed in Table 1. The pictorial representation of H-bond acceptors and H-bond donors are given in Fig. 1. Osiris toxicity analysis suggested no major side effects, signifying their drug ability. A drug score of with yellow color code is considerable as an acceptable drug molecule as given in Table 2.

Auto dock results of Ligand-Protein docking for Aloin and 10 chosen drug target proteins are listed in Table 3, in the order of their binding

energy requirement. Similarly, Auto dock results of Ligand-Protein docking for Aloin and 10 chosen drug target proteins are listed in Table 4, in the order of their binding energy requirement.

Aloin showed the highest affinity toward *P. falciparum* calcium-dependent protein kinase (PfCDPK-2) with binding energy of

**Table 1: Ligand scout analysis of Aloin and microdantin**

Parameters	Aloin	Microdantin
Formula	C <sub>21</sub> H <sub>24</sub> O <sub>9</sub>	C <sub>30</sub> H <sub>32</sub> O <sub>11</sub>
Mol. weight	420.41 Da	568.57 Da
cLogP	-1.536	0.354
TPSA	171.07	197.37
HBA	9	10
HBD	8	8

HBA: Hydrogen bond acceptor; HBD: Hydrogen bond donor; AR: Aromatic ring, TPSA: Topological polar surface area

**Table 2: Osiris molecular property analyzer drug score for Aloin and microdantin**

Parameters	Aloin	Microdantin
Score for cLogP	0.997	0.983
Score for LogS	0.89	0.608
Score for Mol.Wt	0.727	0.316
Score for drug likeness	0.089	0.338
No risk for mutagenicity	1	1
No risk for tumorigenicity	1	1
No risk of irritating effects	1	1
No risk reproductive effects	1	1
Total drug score	0.444 (yellow)	0.351 (yellow)

**Table 3: AutoDock results of Aloin with chosen target proteins**

Target protein	Binding energy	Inhibition constant	Number of H-bonds
PfCDPK2	-8.01	1.35 μM	6
Thioredoxin-2	-7.49	3.22 μM	7
Glutaredoxin-1	-7.11	6.19 μM	6
PfPMT	-7.00	7.35 μM	6
PfPK5	-6.76	11.09 μM	6
Plasmepsin-2	-6.18	29.36 μM	8
PfLDH	-5.97	42.01 μM	4
Falcipain-2	-5.91	46.50 μM	6
PfARF1	-5.58	81.87 μM	6
Pfg27	-5.23	146.48 μM	5

PfCDPK-2: *Plasmodium falciparum* calcium-dependent protein kinases, PfPMT: *Plasmodium falciparum* phosphoethanolamine, PfLDH: *Plasmodium falciparum* lactate dehydrogenase, PfPK-5: *Plasmodium falciparum* protein kinase 5

**Table 4: AutoDock results of microdantin with chosen target proteins**

Target protein	Binding energy	Inhibition constant	Number of H-bonds
Glutaredoxin-1	-8.04	1.28 μM	3
Plasmepsin-2	-7.92	1.57 μM	4
Thioredoxin-2	-7.70	2.25 μM	5
PfPK5	-7.50	3.17 μM	5
PfLDH	-7.27	4.69 μM	7
PfPMT	-6.47	18.13 μM	1
PfCDPK2	-6.42	19.61 μM	4
Falcipain-2	-6.00	39.92 μM	5
PfARF1	-4.78	312.08 μM	2
Pfg27	-3.32	3.7 mM	3

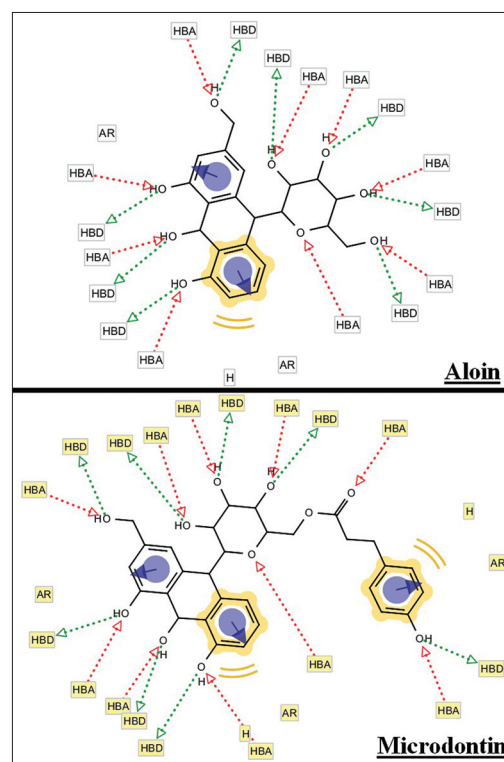
PfCDPK-2: *Plasmodium falciparum* calcium-dependent protein kinases, PfPMT: *Plasmodium falciparum* phosphoethanolamine, PfLDH: *Plasmodium falciparum* lactate dehydrogenase, PfPK-5: *Plasmodium falciparum* protein kinase 5

-8.01 Kcal/Mol and with IC<sub>50</sub> value of 1.35 μM. The PyMol analysis of Aloin and PfCDPK-2 complex is given in Fig. 2. Aloin formed 6 hydrogen bonds with 4 amino acid residues, Ile-212 (2.0 Å), Cys-149 (1.9 Å, 2.5 Å and 2.1 Å), Gly-151 (2.2 Å) and Glu-153 (2.1 Å).

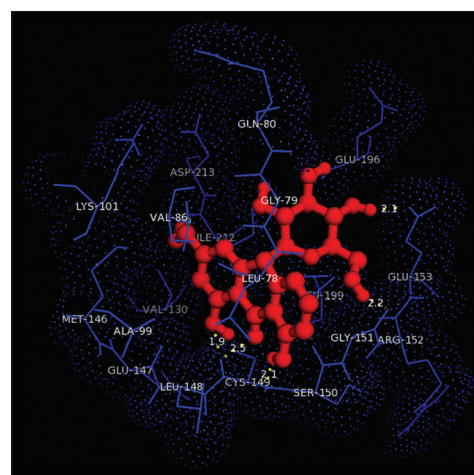
Microdantin showed the highest affinity toward Glutaredoxin-1 with a binding energy of -8.04 Kcal/Mol and IC<sub>50</sub> value of 1.28 μM. LigPlot + analysis of microdantin and glutaredoxin-1 is given in Fig. 3. microdantin formed 3 hydrogen with two amino acid residues, Lys-82 (2.96 Å) and Leu-69 (2.90 Å and 2.76 Å).

## DISCUSSION

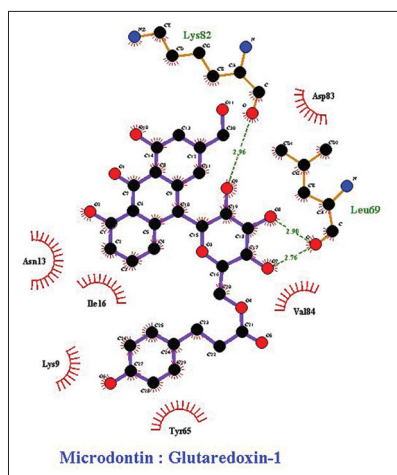
Ligand scout and osiris toxicity analysis revealed the physiochemical properties of the ligand molecules. Both Aloin and microdantin



**Fig. 1: H-bond acceptor and donor sites in Aloin and microdantin, (HBA: Hydrogen bond acceptor, HBD: Hydrogen bond donor, AR: Aromatic ring)**



**Fig. 2: PyMol view of Aloin and *Plasmodium falciparum* calcium-dependent protein kinases-2 complex**



**Fig. 3: LigPlot + analysis of microdointin and glutaredoxin-1 complex**

contains a high number of H-Bond Acceptors and H-Bond Donors. Osiris toxicity analysis displayed that, both Aloin and microdointin possess no major side effects and has significant properties for being used as drug. The report by Gerzihier *et al.*, 2014, demonstrated that Aloin and microdointin did not display any adverse side effects and toxicity, with a maximum of 2 g/kg body weight of pure compound for up to 14 days in mice models. Though the anti-malarial activity of Aloin and microdointin have been proven *in-vitro* and *in-vivo* (Gerzihier *et al.*, 2014), the mechanism by which these compounds exerts its action is still unclear. It was also reported by Gerzihier *et al.*, 2014, that Aloin and microdointin displayed reduced activity individually when compared to crude latex.

Auto dock results along with visual confirmation have led to the suggestion of possible target proteins of the test molecules. Aloin showed the highest significance of inhibition toward PfCDPK-2 (RCSB-PDB ID: 4MVF). Aloin demonstrated 1.35  $\mu\text{M}$  inhibition constant with -8.01 Kcal/Mol binding energy. Since Aloin is rich in hydroxyl group, it has formed 6 hydrogen bonds with PfCDPK-2. This suggests PfCDPK-2 could be the potential target protein for Aloin to exert its antimalarial activity. CDPKs are one of the most common drug targets used for malaria and other protozoan diseases since CDPKs does not exist in mammals. Microdointin on the other hand demonstrated highest significant inhibition toward glutaredoxin-1 (RCSB-PDB ID: 4M2C). It is a well-known drug target and Grx-1 is a key redox protein present in *Plasmodium*. Microdointin showed 1.28  $\mu\text{M}$  inhibition constant with -8.04 Kcal/Mol free binding energy and formed 3 hydrogen bonds.

## CONCLUSION

The objective of this study was achieved by suggesting the potential target protein for Aloin and microdointin for its anti-malarial activity. PfCDPK-2 has been suggestively concluded as the possible drug target for Aloin. CDPK family proteins are a novel drug targets for plasmodium diseases since CDPKs does not exist in mammals.

Glutaredoxin-1 has been suggestively concluded as the potential drug target for microdointin. Since Grx-1 plays key role in the protection of plasmodium cells against Reactive-Oxygen-Species, inhibiting these proteins would lead to the destruction of plasmodium cells by reactive oxygen species. Since Gerzihier *et al.*, 2014 has reported that Aloin and microdointin showed reduced anti-malarial activity when administered individually, it could be suggested that, these two molecules work synergistically together by inhibiting PfCDPK-2 and Grx-1, respectively.

## REFERENCES

1. Kanika P, Dinesh KP. Medicinal importance, pharmacological activities, and analytical aspects of Aloin: A concise report. *J Acute Dis* 2013;2(4):262-3.
2. Gerzihier G, Daniel B, Kaleab A. Isolation, characterization and *in vivo* antimalarial evaluation of anthrones from the leaf latex of *Aloe percrassa* Todaro. *J Nat Rem* 2014;14(2):119-26.
3. World Health Organization. World Malaria Report 2014. United Kingdom Houses of Parliament on 9 December 2014.
4. Maulana T, Hari P. Tea leaves extracted as anti-malaria based on molecular docking plants. *Proc Environ Sci* 2013;17:188-94.
5. Silva AM, Lee AY, Gulnik SV, Maier P, Collins J, Bhat TN, *et al.* Structure and inhibition of plasmepsin II, a hemoglobin-degrading enzyme from *Plasmodium falciparum*. *Proc Natl Acad Sci U S A* 1996;93(19):10034-9.
6. Sharma A, Sharma A, Dixit S, Sharma A. Structural insights into thioredoxin-2: A component of malaria parasite protein secretion machinery. *Sci Rep* 2011;1:179.
7. Yogavel M, Tripathi T, Gupta A, Banday MM, Rahlfs S, Becker K, *et al.* Atomic resolution crystal structure of glutaredoxin 1 from *Plasmodium falciparum* and comparison with other glutaredoxins. *Acta Crystallogr D Biol Crystallogr* 2014;70:91-100.
8. Holton S, Merckx A, Burgess D, Doerig C, Noble M, Endicott J. Structures of *P. falciparum* PfPK5 test the CDK regulation paradigm and suggest mechanisms of small molecule inhibition. *Structure* 2003;11(11):1329-37.
9. Hogg T, Nagarajan K, Herzberg S, Chen L, Shen X, Jiang H, *et al.* Structural and functional characterization of Falcipain-2, a hemoglobinase from the malarial parasite *Plasmodium falciparum*. *J Biol Chem* 2006;281(35):25425-37.
10. Lee SG, Kim Y, Alpert TD, Nagata A, Jez JM. Structure and reaction mechanism of phosphoethanolamine methyltransferase from the malaria parasite *Plasmodium falciparum*: An antiparasitic drug target. *J Biol Chem* 2012;287:1426-34.
11. Cook WJ, Smith CD, Senkovich O, Holder AA, Chattopadhyay D. Structure of *Plasmodium falciparum* ADP-ribosylation factor 1. *Acta Crystallogr Sect F Struct Biol Cryst Commun* 2010;66:1426-31.
12. Sharma A, Sharma I, Kogkasuriyachai D, Kumar N. Structure of a gametocyte protein essential for sexual development in *Plasmodium falciparum*. *Nat Struct Biol* 2003;10(3):197-203.
13. Arrigoni A, Bertini L, De Gioia L, Papaleo E. Inhibitors of the Cdc34 acidic loop: A computational investigation integrating molecular dynamics, virtual screening and docking approaches. *FEBS Open Bio* 2014;4:473-84.
14. Arun KV, Keshav M. *In silico* analysis of indoles against 1KE8 inhibitors using auto dock. *Br J Pharm Res* 2013;3(3):446-53.
15. Jayasree G, Sruthi S. Molecular docking studies of antidiabetic activity of cinnamon compounds. *Asian J Pharm Clin Res* 2014;7(2):31-4.
16. Ingale AG, Goto S. Prediction of CTL epitope, *in silico* modeling and functional analysis of cytolethal distending toxin (CDT) protein of *Campylobacter jejuni*. *BMC Res Notes* 2014;7:92.
17. Dhara L, Tripathi A, Pal A. Molecular characterization and *in silico* analysis of naturally occurring TEM beta-lactamase variants among pathogenic Enterobacteriaceae infecting Indian patients. *Biomed Res Int* 2013;2013:783540.
18. Vaibhav M, Nidhi M, Amrendra NP. Molecular docking studies of anti-HIV drug BMS-488083 derivatives using HEX and GP120 interaction analysis using Pymol. *Int J Sci Res Pub* 2013;3(6):1-7.
19. Seeliger D, de Groot BL. Ligand docking and binding site analysis with PyMOL and Autodock/Vina. *J Comput Aided Mol Des* 2010;24:417-22.
20. Cleave SS, Panda R, Suresh PK. *In silico* exploration of phenytoin binding site in two catalytic states of human P-glycoprotein models. *Indian J Biochem Biophys* 2013;50:7-13.
21. Muthukumaran S, Anusha K, Umashankar V, Sulochana KN, Umashankar V. Elucidation of iron binding patterns through *in silico* approaches in human iron binding proteins. *Int J Ther Appl* 2013;10:11-8.
22. Temml V, Kuehn S, Schuster D, Schwaiger S, Stuppner H, Fuchs D. Interaction of *Carthamus tinctorius* lignan arctigenin with the binding site of tryptophan-degrading enzyme indoleamine 2,3-dioxygenase. *FEBS Open Bio* 2013;3:450-2.
23. Grienke U, Mihály-Bison J, Schuster D, Afonyushkin T, Binder M, Guan SH, *et al.* Pharmacophore-based discovery of FXR-agonists. Part II: Identification of bioactive triterpenes from *Ganoderma lucidum*. *Bioorg Med Chem* 2011;19:6779-91.