

## REVIEW OF MEDICINAL USES, PHYTOCHEMISTRY, AND PHARMACOLOGICAL PROPERTIES OF *DRIMIA ELATA*

ALFRED MAROYI\*

Department of Botany, Medicinal Plants and Economic Development Research Centre, University of Fort Hare, Private Bag X1314, Alice, South Africa. Phone: 0027719600326. Email: amaroyi@ufh.ac.za

Received: 18 December 2018, Revised and Accepted: 11 February 2019

**ABSTRACT**

*Drimia elata* is an important and well-known medicinal plant in tropical Africa. This study critically reviewed the medicinal applications, phytochemistry, and pharmacological activities of *D. elata*. Literature on medicinal applications, phytochemical, and pharmacological activities of *D. elata* was collected from multiple internet sources including Elsevier, Google Scholar, SciFinder, Web of Science, PubMed, BMC, ScienceDirect, and Scopus. Complementary information was gathered from pre-electronic sources such as books, book chapters, theses, scientific reports, and journal articles obtained from the university library. This study showed that *D. elata* is used for treating several medical conditions, particularly general ailments, blood and cardiovascular system, reproductive system and sexual health, urinary system, infections and infestations, digestive system, respiratory system, and muscular-skeletal system disorders. Phytochemical compounds identified from the species include bufadienolides, alkaloids, aromatic acids, flavonoids, phlobatannins, saponins, steroids, tannins, and terpenoids. Ethnopharmacological research revealed that *D. elata* extracts have acetylcholinesterase enzyme inhibitory, antibacterial, antifungal, antimycobacterial, anticancer, anti-inflammatory, antioxidant, hemagglutinating, and cytotoxicity activities. *D. elata* should be subjected to extensive *in vivo* experiments and also future studies should focus on how potential toxic components of the species can be managed when it is used as herbal medicine.

**Keywords:** Asparagaceae, *Drimia elata*, Herbal medicine, Tropical africa.

© 2019 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>) DOI: <http://dx.doi.org/10.22159/ajpcr.2019.v12i4.30963>

**INTRODUCTION**

*Drimia elata* Jacq. is an important and well-known medicinal plant in South Africa. Van *et al.* [1] provide an excellent introduction to the ethnopharmacological properties of *D. elata* and several other important medicinal plants in South Africa. *D. elata* is an ingredient of at least two traditional herbal concoctions in South Africa, known as “*imbizae phuzwato*” and “*intelezi*” that are sold commercially in the country. A herbal tonic, *imbizae phuzwato* is made from a mixture of roots, bulbs, rhizomes, and leaves of *Acokanthera oppositifolia* (Lam.) Codd, *Aster bakeranus* Burt Davy ex C.A. Sim., *Corchorus asplenifolius* Burch., *Cyrtanthus obliquus* (L.f.) Aiton, *Fusifilum physodes* (Jacq.) Raf. ex Speta, *Eriosema cordatum* E.Mey., *Gnidia kraussiana* Meisn. var. *kraussiana*, *Gomphocarpus fruticosus* (L.) W.T. Aiton, *Gunnera perpensa* L., *Hypericum aethiopicum* Thunb., *Ledebouria* spp., *Lycopodium clavatum* L., *Momordica balsamina* L., *Rubia cordifolia* L., *Scadoxus puniceus* (L.) Friis and Nordal, *Stephania abyssinica* (Quart.-Dill. and A. Rich.) Walp., *Tetradenia riparia* (Hochst.) Codd, *Vitellariopsis marginata* (N.E.Br.) Aubrév, *Watsonia densiflora* Bak., and *Zanthoxylum capense* (Thunb.) Harv. [2,3]. The concoction is used as an energizing and detoxifying tonic used against general body pains, stress, constipation, arthritis, kidney problems, high blood pressure, and to increase sexual prowess [2,3]. *D. elata* is also an ingredient of “*intelezi*,” whose plant species composition varies from region to region of South Africa. *Intelezi* is used to protect households from evil spirits and lightning, and also to chase away, ward off or root out evil spirits [4].

*D. elata* is the third most popular bulbous medicinal plant used in South African traditional therapy [5] and is one of the topmost wild-harvested species sold in the informal economy trade in the Eastern Cape [6,7], Gauteng [8-10], KwaZulu-Natal [8,11], Limpopo [12], and the Western Cape [7,13,14] provinces in South Africa. Research by Ndawonde *et al.* [15] showed that *D. elata* bulb was sold by >50.0% of the traders in KwaZulu-Natal Province, while Philander *et al.* [14] revealed that bulbs of the species were sold by 35% of the traders in the Western

Cape province, with 60.48 kg of the bulb fetching US\$26.21. Earlier research by Dold and Cocks [6] revealed that *D. elata* is among the most frequently traded species in the Eastern Cape Province with 113.9 kg as the mean quantity traded per trader per annum with a kilogram of the bulb fetching US\$3.36. Marshall [16] argued that *D. elata* is scarce and heavily traded in South Africa, characterized by a high monetary value in the country. Due to increasing demand for the species, *D. elata* is managed in herbal medicine home gardens in the Eastern Cape [17], Limpopo [18-20], and the Western Cape [14] provinces. Research by Wiersum *et al.* [17] revealed that *D. elata* is among the ten most frequently cultivated herbal medicines in medicinal home gardens in the Eastern Cape Province. It is, therefore, within this context that the current study was undertaken aimed at summarizing the medicinal uses, phytochemical, and ethnopharmacological properties of *D. elata* so as to evaluate its therapeutic importance throughout its distributional range.

**BOTANICAL PROFILE AND DESCRIPTION OF *D. ELATA***

The genus *Drimia* Jacq. is a large group of deciduous geophytes belonging to the family Asparagaceae, previously included in the Hyacinthaceae family. The family of Hyacinthaceae is divided into four monophyletic subfamilies, namely Hyacinthoideae, Ornithogaloideae, Ozirioideae, and Urginoideae [21]. At present, this family is considered as a subfamily Scilloideae in the expanded Asparagaceae sensu [22,23]. The species in each subfamily synthesize specialized secondary metabolites with Hyacinthoideae synthesizing homoisoflavanones and triterpenoids, Ornithogaloideae (cardenolides and steroidal glycosides), and Urginoideae synthesizing bufadienolides [21]. The subfamily Urginoideae has flat or winged seeds characterized by brittle, loosely adhering test comprising genera *Bowiea* Harv. ex Hook. f. and *Drimia* [24]. The genus *Drimia* is described by Manning *et al.* [24,25] in an inclusive and broad sense, including genera such as *Litanthus* Harv., *Mucinea* M. Pinter *et al.*, *Rhadamanthus* Salisb., *Rhodocodon* Baker, *Sagittanthera* Mart-Azorín *et al.*, *Thuranthos* C. H. Wright, *Tenicroa* Raf., and *Urginea* Steinh. The taxonomy of genus *Drimia* has always

been difficult with several species treated under genus *Urginea* until Jessop [26] reduced *Urginea* to a synonym of *Drimia*. The genus consists of about 100 bulbous species distributed in Southern Africa through tropical Africa to the Mediterranean, Asia, and Madagascar [24]. Synonyms of *D. elata* include *D. alta* R.A. Dyer, *D. ciliaris* Jacq. ex Willd., *D. purpurascens* J. Jacq., *D. robusta* Baker, *D. villosa* (Lindl.) Kunth, *D. zombensis* Baker, *Idotheaelata* Kunth, *I. ciliaris* (Jacq. ex Willd.) Kunth, *I. purpurascens* (J. Jacq.) Kunth, *I. robusta* (Baker) Kuntze, and *I. villosa* (Lindl.) Kunth [1,27-31].

*D. elata* is a geophyte with large underground bulb, strap-shaped leaves and long, slender flowering stalk which grows up 1.8 m in height [1,27-29,31]. The flowers are tubular, whitish to purple in color with the tips of the petals characteristically reflexed and the stamens fused into a narrow tube [1]. *D. elata* has been recorded in grassland, often among rocks at an altitude ranging from 15 m to 1650 m above sea level [29]. The species has been recorded in Botswana, Angola, Malawi, Kenya, South Africa, Swaziland, Zambia, South Sudan, Tanzania, Sudan, Uganda, Zimbabwe, and Mozambique [1,27-31](Fig. 1).

### MEDICINAL USES OF *D. ELATA*

The medicinal applications recorded from literature were classified into ten medical categories following the Economic Botany Data Collection Standard [32] with some changes proposed by Macía *et al.* [33] and Gruca *et al.* [34]. This review showed that *D. elata* is used for treating several medicinal conditions, particularly general ailments, blood and cardiovascular system, reproductive system and sexual health, urinary system, infections and infestations, digestive system, respiratory system, and muscular-skeletal system disorders (Fig. 2). *D. elata* is used as herbal medicine against three out of five diseases categorized by the World Health Organization (WHO) as the top five killer diseases in sub-Saharan Africa in 2012. These diseases include human immunodeficiency virus/acquired immune deficiency syndrome (HIV/AIDS), lower respiratory tract infections, and diarrheal diseases [35]. Most medicinal uses are linked to the bulb and leaf or the entire plant in ritual or magical uses, and the species is also used mixed with other plant species (Table 1). Research by Gurib-Fakim [36] and Maroyi [37,38] revealed that traditional medicines are often prepared by combining several different plant species to effect synergistic properties or to initiate an interaction with a relevant molecular target.

### PHYTOCHEMICAL CONSTITUENTS OF *D. ELATA*

*D. elata* is characterized by cardiac glycosides, particularly bufadienolides. All the bufadienolides that have been isolated from *D. elata* are collated in Table 2. Kellerman *et al.* [59,60] argued that bufadienolide containing plants are toxic to livestock with an estimated 33% of plant-related mortality in cattle in South Africa attributed to this compound. Van *et al.* [61] argued that there is a danger of accidental poisoning or that people may be harmed if bulbs of *D. elata* are used indiscriminately as rubbing the bulb scales or leaves on bare skin produces a stinging and irritating effect and a skin rash is produced. However, bufadienolides are known to have a wide range of biological activities including anti-tumor, antiproliferative, and cytotoxic activities [62-68].

Koorbanally *et al.* [69] identified aromatic acids, 4-hydroxy-3-methoxybenzoic acid, 3,4-dihydroxybenzoic acid, and trans-3-(4'-hydroxyphenyl)-2-propenoic acid from the ethyl acetate bulb extract of *D. elata*. Matotoka and Masoko [70] identified flavonoids, phlobatannins, saponins, tannins, and terpenoids from the *D. elata* bulb (Table 3). Similarly, Matotoka and Masoko [41] identified alkaloids, flavonoids, saponins, steroids, tannins, and terpenoids from a herbal mixture of *D. elata* bulb mixed with leaves of *Monsonia angustifolia*, *Sarcostemma viminale* and *Vahlia capensis*, *Kirkia wilmsii* (leaves, roots, and twigs), and *Hypoxis hemerocallidea* (corm).

Okem *et al.* [71] argued that *D. elata* bulbs obtained from the herbal medicine informal markets in Pietermaritzburg, KwaZulu-Natal Province in South Africa contained high levels of heavy metals, with



Fig. 1: Distribution of *Drimia elata* in tropical Africa

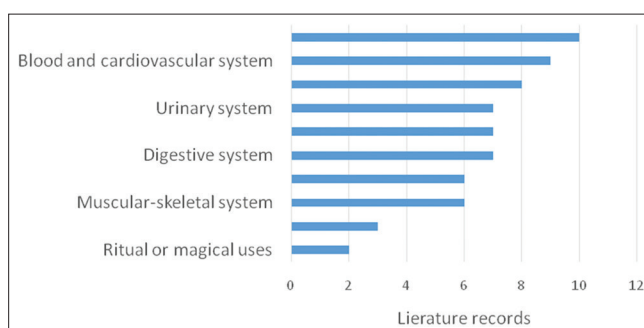


Fig. 2: Number of literature records per medicinal category of *Drimia elata* following the economic botany data collection standard [32]

aluminum, cadmium, manganese, and chromium being above the WHO recommended safety levels (Table 3). Quantities of mineral elements and phytochemical compounds isolated from *D. elata* are listed in Table 3.

### PHARMACOLOGICAL PROPERTIES OF *D. ELATA*

Pharmacological studies on *D. elata* bulb and leaf extracts exhibited potent *in vitro* pharmacological activities including acetylcholinesterase enzyme inhibitory [2], antibacterial [2,43,50,70-80], antifungal [2,76], antimycobacterial [50], anticancer [81], anti-inflammatory [2,72,82,83], antioxidant [43,70], hemagglutinating [84], and cytotoxicity [43] activities.

#### Acetylcholinesterase enzyme inhibitory activities

Ndhala *et al.* [2] investigated the acetylcholinesterase enzyme inhibitory activities of aqueous bulb extracts of *D. elata* using the enzyme isolated from electric eels with galanthamine as the positive control. The extract showed moderate AChE inhibitory activity of 50.0% with half maximal inhibitory concentration ( $IC_{50}$ ) value of  $487.4 \pm 8.0 \mu\text{g/mL}$  [2]. The ability of *D. elata* bulb extracts to inhibit acetylcholinesterase shows potential therapeutic potential of the species in the management of memory loss and neurodegenerative disorders.

#### Antibacterial activities

Luyt *et al.* [72] evaluated antibacterial activities of aqueous, ethyl acetate, and ethanol bulb and leaf extracts of *D. elata* against *Bacillus*

Table 1: Medicinal uses of *D. elata*

Medicinal use	Parts of the plant used	References
Blood and cardiovascular system		
Blood purification	Bulbs or <i>imbizae phuzwata</i> concoction	[2,3,39-42]
Blood purification	Bulbs mixed with leaves, roots, and twigs of <i>Kirkia wilmsii</i> Engl., <i>Hypoxis hemerocallidea</i> Fisch., C. A. Mey. and Avé-Lall. (corms), <i>Monsonia angustifolia</i> E. Mey. ex A. Rich. (leaves), and leaves of <i>Sarcostem maviminale</i> (L.) Br. and <i>Vahlia capensis</i> (L. f.) Thunb.	[43]
Hypertension	Bulbs	[2,3,19,44]
Digestive system		
Antidiarrheal	Bulbs mixed with leaves, roots, and twigs of <i>Kirkia wilmsii</i> , <i>Hypoxis hemerocallidea</i> (corms), <i>Monsonia angustifolia</i> (leaves), and leaves of <i>Sarcostem maviminale</i> , and <i>Vahlia capensis</i>	[43]
Constipation	<i>Imbizae phuzwata</i> concoction	[2,3]
Emetic	Bulbs	[37,45-47]
General ailments		
Angina pain	Bulbs	[48]
Body pains	<i>Imbizae phuzwata</i> concoction	[2,3]
Energizing tonic	<i>Imbizae phuzwata</i> concoction	[2,3]
Fever	Bulbs and leaves or bulbs mixed with roots of <i>Artemisia afra</i> Jacq. ex Willd., <i>Siphonochilus aethiopicus</i> (Schweinf.) B. L. Burt and <i>Erythrina caffra</i> Thunb.	[45,49,50]
Headache	Bulbs	[1]
Heart tonic	Bulbs	[47]
Internal sores	Bulbs	[41]
Sores	Bulbs and leaves	[51]
Stress	<i>Imbizae phuzwata</i> concoctions	[2,3]
Infections and infestations		
Gonorrhea	Bulbs	[19,52,53]
HIV/AIDS opportunistic infections	Bulbs or bulbs mixed with twigs of <i>Sarcostem aviminale</i> (L.) R. Br. and roots of <i>Elaeodendron transvaalense</i> (Burt Davy) R. H. Archer, <i>Elephantor rhizaelephantina</i> (Burch.) Skeels and <i>Zanthoxylum capense</i> (Thunb.) Harv. and bark of <i>Sclero caryabirrea</i> (A. Rich.) Hochst.	[19,53,54]
STIs	Bulbs mixed with roots of <i>Elaeodendron transvaalense</i> , <i>Elephantor rhizaelephantina</i> (roots), <i>Sarcostem aviminale</i> (twigs), <i>Scleroc aryabirrea</i> (bark), and <i>Zanthoxylum capense</i> (root)	[55]
Tuberculosis	Bulbs or bulbs mixed with roots of <i>Callilep islaureola</i> DC, <i>Croton menyharthii</i> Pax, <i>Senna italica</i> Mill. and bulb of <i>Siphonochilus aethiopicus</i> (Schweinf.) B. L. Burt or bulbs mixed with the bark of <i>Warburgi asalutaris</i> (G. Bertol.) Chiov. or bulbs mixed with leaves of <i>Ricinus communis</i> L. or bulbs mixed with roots of <i>Dicoma anomala</i> Sond. and bulb of <i>Eucomis autumnalis</i> (Mill.) Chitt.	[48,50]
Muscular-skeletal system		
Arthritis	Bulbs and <i>imbizae phuzwata</i> concoction	[2,3,39]
Back pain	Bulbs	[41]
Edema	Bulbs	[1]
Inflammation	Bulbs and leaves	[51]
Muscle pain	Bulbs	[41]
Swelling	Bulbs	[39]
Pain		
Pain	Bulbs and leaves or bulb mixed with leaves, roots, and twigs of <i>Kirkia wilmsii</i> , <i>Hypoxis hemerocallidea</i> (corm), <i>Monsonia angustifolia</i> (leaves), and leaves of <i>Sarcostemma viminale</i> and <i>Vahlia capensis</i>	[43,46,51]
Reproductive system and sexual health		
Aphrodisiac	<i>Imbizae phuzwata</i> concoction or bulbs mixed with leaves, roots, and twigs of <i>Kirkia wilmsii</i> , <i>Hypoxis hemerocallidea</i> (corm), <i>Monsonia angustifolia</i> (leaves), and leaves of <i>Sarcostemma viminale</i> and <i>Vahlia capensis</i>	[2,3,43]
Erectile dysfunction	Bulbs	[56]
Impotence	Bulbs	[19,57]
Infertility	Bulbs	[1,18,19,57]
Respiratory system		

(Contd...)

Table 1: (Continued)

Medicinal use	Parts of the plant used	References
Blocked nose	Bulbs or bulbs mixed with roots of <i>Artemisia afra</i> , <i>Siphonochilus aethiopicus</i> , and <i>Erythrina caffra</i>	[50]
Chest pains	Bulbs and leaves or bulbs mixed with leaves of <i>Lippia javanica</i> (Burm. f.) Spreng. or bulbs mixed with roots of <i>Artemisia afra</i> , <i>Siphonochilus aethiopicus</i> , and <i>Erythrina caffra</i> or bulbs mixed with roots of <i>Elaeodendron transvaalense</i> (Burt Davy) R. H. Archer	[48-50]
Colds	Bulbs and leaves	[45,46,49]
Cough	Bulbs mixed with leaves of <i>Lippia javanica</i>	[50]
Expectorant	Bulbs	[45,47]
Runny nose	Bulbs mixed with leaves of <i>Lippia javanica</i>	[50]
Ritual or magical uses		
Protect households from evil spirits and lightning, and also to chase away, ward off or root out evil spirits	<i>Intelezi</i> herbal concoction	[4,45]
Urinary system		
Bladder complaints	Leaves	[1,15,58]
Diuretic	Leaves	[1,39,47,58]
Kidney problems	<i>Imbizae phuzwata</i> concoction	[2,3]
Uterus problems	Bulbs and leaves	[1,15,58]

HIV: Human immunodeficiency virus, AIDS: Acquired immune deficiency syndrome, STIs: Sexually transmitted infections

Table 2: Bufadienolides isolated from *D. elata* bulb using NMR spectroscopy

Bufadienolides	Extract	References
Proscillaridin A	Chloroform: isopropanol	[72,73]
Scilliroside	Chloroform or chloroform-n-butanol	[74]
12 $\beta$ -hydroxyscillirosidin	Chloroform or chloroform-n-butanol	[74]
12 $\beta$ -hydroxyscilliroside	Chloroform or chloroform-n-butanol	[74]
Hellebrigenin-3-0- $\beta$ -glucoside	Chloroform or chloroform-n-butanol	[74]
16 $\beta$ -hydroxyhellebrigenin	Chloroform or chloroform-n-butanol	[74]
16 $\beta$ -hydroxyhellebrigenin-3-0- $\beta$ -glucoside	Chloroform or chloroform-n-butanol	[74]
5 $\beta$ ,16 $\beta$ -dihydroxybufalin-3-0- $\beta$ -glucoside	Chloroform or chloroform-n-butanol	[74]
6 $\beta$ -acetoxy-3 $\beta$ ,8 $\beta$ ,12 $\beta$ ,14 $\beta$ -tetrahydroxybufa-4,20,22-trienolide (12 $\beta$ -hydroxyscillirosidin)	Dichloromethane	[75]
14 $\beta$ -hydroxybufa-4,20,22-trienolide 3 $\beta$ -O-( $\alpha$ -L-rhamnopyranosyl)-(1 $\rightarrow$ 4)- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)- $\alpha$ -L-rhamnopyranoside} (arginin)	Dichloromethane	[75]
6 $\beta$ -acetoxy-3 $\beta$ ,8 $\beta$ ,14 $\beta$ -trihydroxy-12-oxobufa-4,20,22-trienolide	Dichloromethane	[69]
6 $\beta$ -acetoxy-3 $\beta$ ,8 $\beta$ ,12 $\beta$ ,14 $\beta$ -tetrahydroxybufa-4,20,22-trienolide (12 $\beta$ -hydroxyscillirosidin)	Dichloromethane	[69]

NMR: Nuclear magnetic resonance

*subtilis*, *Escherichia coli*, *Klebsiella pneumoniae*, *Micrococcus luteus*, *Pseudomonas aeruginosa*, *Staphylococcus aureus*, and *Staphylococcus epidermidis* using disk-diffusion assay with neomycin (2  $\mu$ g/ml) as the positive control. Only ethyl acetate bulb extract was active against *B. subtilis*, *K. pneumoniae*, *M. luteus*, *P. aeruginosa*, and *S. aureus* with inhibition ratios ranging from 0.1 to 0.63 [72]. Ncube *et al.* [76] evaluated antibacterial activities of aqueous, dichloromethane, ethanol, and petroleum ether extracts of bulb and leaves of *D. elata* between spring, summer, autumn, and winter seasons against *Bacillus subtilis*, *S. aureus*, *E. coli*, and *K. pneumoniae* using the microdilution bioassay with neomycin ( $\mu$ g/ml) as the positive control. The extracts were active in all seasons except for winter when the leaves are not available showing minimum inhibitory concentration (MIC) values ranging from 0.8 mg/ml to >12.5 mg/ml [76]. Ndhala *et al.* [2] evaluated the antibacterial activities of aqueous, petroleum ether, dichloromethane, and ethanol bulb extracts of *D. elata* against *Bacillus subtilis*, *E. coli*, *K. pneumoniae*, and *S. aureus* using the microdilution bioassay with neomycin as the positive control. The extracts showed activities with MIC values ranging from 0.8 to >12.5 mg/ml [2]. Baskaran *et al.* [77] evaluated the antibacterial activities of ethanol bulb, leaf, shoots, and plantlet extracts of *in vitro* and *ex vitro* regenerated *D. elata* in comparison to naturally-grown plants against *S. aureus*, *Enterococcus faecalis*, *E. coli*, and *P. aeruginosa* using the microdilution method with neomycin (100  $\mu$ l) as the positive control. All extracts exhibited activities with MIC values ranging from 0.2 mg/ml to 12.5 mg/ml [77]. Okem *et al.* [71] evaluated antibacterial activities of ethanol stem bulb extracts of *D. elata* against *E. coli* and *S. aureus* using microdilution assay with neomycin (2  $\mu$ g/ml)

as the positive control. The extracts exhibited activities with MIC values ranging from 6.3 mg/mL to 12.5 mg/mL [71]. Okem *et al.* [78] evaluated the effects of cadmium and aluminum accumulation on antibacterial activities of ethanol stem bulb extracts of *D. elata* against *E. coli* and *S. aureus* using microdilution assay with neomycin (2  $\mu$ g/ml) as the positive control. The control extracts exhibited MIC values of 0.4 mg/ml and 0.8 mg/ml against *S. aureus* and *E. coli*, respectively, while antibacterial activities decreased in extracts exposed to increasing heavy metal stress with MIC values ranging from 0.8 mg/ml to 12.5 mg/ml [78]. Madisha [50] evaluated the antibacterial activities of ethanol, methanol, hydroethanol, and dichloromethane bulb extracts of *D. elata* against *Bacillus cereus*, *E. faecalis*, *E. coli*, *Neisseria gonorrhoeae*, *Proteus vulgaris*, *P. aeruginosa*, *Shigella flexneri*, *S. aureus*, *Staphylococcus epidermidis*, and *Vibrio parahaemolyticus* using agar well dilution method and streak plate disc diffusion assays. The extracts revealed varying degrees of activities with the zone of inhibition values ranging from 8.0 mm to 19.0 mm and MIC values ranging from 0.1 mg/mL to 12.5 mg/mL [50]. Madisha [50] also evaluated the antibacterial activities of ethanol and hydroethanol bulb extracts of *D. elata* mixed with roots of *Elephantorrhiza elephantina* and leaves of *Aloe marlothii* and *Maurea angolensis* against *B. cereus*, *E. faecalis*, *E. coli*, *N. gonorrhoea*, *P. vulgaris*, *P. aeruginosa*, *S. flexneri*, *S. aureus*, *S. epidermidis*, and *V. parahaemolyticus* using agar well dilution method and streak plate disk diffusion assays. The extracts exhibited activities against tested pathogens with MIC values ranging from 0.4 mg/mL to 1.6 mg/mL [50]. Matotoka and Masoko [70] evaluated antibacterial activities of acetone and hexane extracts of *D. elata* bulb against *S. aureus*, *E. faecalis*, *E. coli*, and *P. aeruginosa* using the broth

Table 3: Mineral and phytochemical composition of *D. elata*

Mineral and phytochemical composition	Values	Plant parts	References
Aluminum (mg/kg dry weight)	559.8–1595.0	Bulbs	[70,71]
Arsenic (mg/kg dry weight)	1.8	Bulbs	[71]
Boron (mg/L)	3.0	Bulbs	[70]
Cadmium (mg/kg dry weight)	0.01–0.06	Bulbs	[71]
Calcium (mg/L)	19.0	Bulbs	[70]
Cobalt (mg/L)	0.04	Bulbs	[70]
Copper (mg/kg dry weight)	5.6–11.3	Bulbs	[71]
Chromium (mg/kg dry weight)	7.8–12.0	Bulbs	[71]
Flavonoid (mg of quercetin equivalent/g extract)	0.54–15.0	Bulbs and leaves	[43,71,76]
Gallotannin ( $\mu$ g gallic acid equivalent/g dry weight)	4.0–7.0	Bulbs and leaves	[76]
Iron (mg/L)	0.15	Bulbs	[70]
Iron (mg/kg dry weight)	593.0–1634.0	Bulbs	[71]
Lead (mg/kg dry weight)	0.2–1.2	Bulbs	[71]
Magnesium (mg/L)	28.0	Bulbs	[70]
Manganese (mg/kg dry weight)	60.7–145.8	Bulbs	[70,71]
Mercury (mg/kg dry weight)	0.04–0.8	Bulbs	[71]
Molybdenum (mg/L)	0.02	Bulbs	[70]
Nickel (mg/kg dry weight)	4.2–10.0	Bulbs	[71]
Phosphorus (mg/L)	24.0	Bulbs	[70]
Potassium (mg/L)	53.0	Bulbs	[70]
Silicon (mg/L)	4.0	Bulbs	[70]
Sodium (mg/L)	56.0	Bulbs	[70]
Sulfur (mg/L)	7.0	Bulbs	[70]
Tannin (mg of gallic acid equivalent/g extract)	4.5–9.6	Bulbs and leaves	[43,76]
Tin (mg/kg dry weight)	31.4–79.8	Bulbs	[71]
Total phenolics (mg gallic acid equivalent/g dry weight)	0.05–2.5	Bulbs and leaves	[43,71,76]
Total saponins (mg diosgenin equivalent/g dry weight)	5.0–17.0	Bulbs and leaves	[76]
Total steroidal saponin (mg diosgenin equivalent/g dry weight)	1.0–4.5	Bulbs and leaves	[76]
Zinc (mg/L)	0.1	Bulbs	[70]
Zinc (mg/kg dry weight)	34.1–102.6	Bulbs	[71]

microdilution assay. The extracts exhibited activities against *E. faecalis* and *P. aeruginosa* with MIC values ranging from 0.6 mg/mL to 2.5 mg/mL and total activities ranging from 3.3 mL/g to 13.3 mL/g [70]. Baskaran *et al.* [79] evaluated antibacterial activities of the aqueous bulb and root extracts of *ex vitro* grown *D. elata* derived from somatic embryogenesis against *Bacillus subtilis*, *E. faecalis*, *M. luteus*, *S. aureus*, *E. coli*, *K. pneumoniae*, and *P. aeruginosa* using microtiter bioassay with neomycin ( $\mu$ g/ml) as a positive control. The extracts exhibited activities with MIC values ranging from 0.4 mg/ml to 6.3 mg/ml [79]. Kandari [80] evaluated antibacterial activities of aqueous, dichloromethane, and ethanol bulb extracts of *D. elata* subjected to vermicompost leachate at different concentrations against *Bacillus subtilis*, *S. aureus*, and *E. coli* using microdilution assay. The extracts exhibited activities with MIC values ranging from 0.4 mg/ml to 6.3 mg/ml [80]. Matotoka and Masoko [43] evaluated antibacterial activities of an herbal mixture of *D. elata* bulb together with leaves of *M. angustifolia*, *S. viminale* and *Vahlia capensis*, *Kirkia wilmsii* (leaves, roots, and twigs), and *Hypoxis hemerocallidea* (corm) against *Bacillus subtilis*, *Citrobacter braakii*, *Enterobacter aerogenes*, *Enterobacter cloacae*, *E. faecalis*, *E. coli*, *K. pneumoniae*, *Leclercia adecarboxylata*, *Pantoea agglomerans*, *P. aeruginosa*, and *S. aureus* using broth microdilution assay with ampicillin ( $\mu$ g/mL) as a positive control. The herbal mixture exhibited activities with MIC values ranging from 0.3 mg/mL to >2.5 mg/mL [43]. The documented antibacterial activities exhibited by extracts of *D. elata* corroborate the traditional application of the species as herbal medicine against bacterial infections causing diarrhea [43], gonorrhoea [19,52,53], sexually transmitted infections [55], and sores [51].

#### Antifungal activities

Ncube *et al.* [76] evaluated antifungal activities of aqueous, dichloromethane, ethanol, and petroleum ether extracts of bulb and leaf extracts of *D. elata* between spring, summer, autumn, and winter seasons against *Candida albicans* using the microdilution bioassay with amphotericin B ( $\mu$ g/ml) as the positive control. The extracts were active in all seasons except for winter when the leaves are not available showing MIC and MFC values ranging from 0.4 mg/ml to

>12.5 mg/ml [76]. Ndhala *et al.* [2] investigated the antifungal activity of aqueous, petroleum ether, dichloromethane, and ethanol bulb extracts of *D. elata* against *C. albicans* using the microdilution assay with amphotericin B as the positive control. The extracts exhibited activities with MIC and MFC values ranging from 3.1 to 6.3 mg/mL and 6.3 mg/mL to 12.5 mg/mL [2].

#### Antimycobacterial activities

Madisha [50] evaluated the antimycobacterial activities of ethanol, methanol, hydroethanol, and dichloromethane bulb extracts of *D. elata* against *Mycobacterium tuberculosis*, *Mycobacterium smegmatis*, *Mycobacterium peregrinum*, and *Mycobacterium haemophilus* using agar well dilution method and streak plate disc diffusion assays. The extracts revealed varying degrees of activities with the zone of inhibition values ranging from 9.0 mm to 21.0 mm and MIC values ranging from 0.1 mg/mL to 12.5 mg/mL [50]. Madisha [50] also evaluated the antimycobacterial activities of ethanol and hydroethanol bulb extracts of *D. elata* mixed with roots of *Elephantor rhizaelephantina* and leaves of *A. marlothii* and *M. angolensis* against *M. tuberculosis*, *M. smegmatis*, *M. peregrinum*, and *M. haemophilus* using agar well dilution method and streak plate disc diffusion assays. The extracts exhibited activities against tested pathogens with MIC values ranging from 0.1 mg/mL to 1.6 mg/mL [50]. These findings show the potential of *D. elata* in the treatment and management of respiratory problems such as blocked nose [48], chest pains [46-48], colds [43,44,47], cough [48], and runny nose [50].

#### Anticancer activities

Fouche *et al.* [81] evaluated *in vitro* anticancer activities of dichloromethane:methane (1:1) of the whole plant of *D. elata* against a panel of three human cell lines (breast MCF7, renal TK10, and melanoma UACC62). The extract exhibited total growth inhibition values ranging from 6.3  $\mu$ g/ml to 29.6  $\mu$ g/ml. The extracts were screened against 60 human cancer cell lines organized into sub-panels representing leukemia, melanoma, cancer of the lung, colon, kidney, ovary, central nervous system, breast, and prostate. The extract exhibited total

growth inhibition values of 1.1 µg/ml against ovarian (OVCAR-3), 1.4 µg/ml against central nervous system cancer, CNSC SF-539 and 1.4 µg/ml against non-small cell lung cancer, NSCLC A549/ATCC [81]. The documented anticancer activities may be attributed to bufadienolides as these compounds are known to have anticancer activities [62-68].

#### Anti-inflammatory activities

Luyt *et al.* [72] evaluated anti-inflammatory activities of aqueous, ethyl acetate, and ethanol bulb and leaf extracts of *D. elata* using the cyclooxygenase assay with indomethacin as the positive control. The bulb extracts inhibited cyclooxygenase with inhibition ranging from 69.0% to 98.0% which was comparable to 94% exhibited by indomethacin, the positive control [72]. Stafford *et al.* [82] evaluated anti-inflammatory activities of aqueous, ethanol, and hexane bulb extracts of fresh and stored material of *D. elata* by assessing their ability to inhibit cyclooxygenase (COX)-1 enzymes. The ethanol extract showed high inhibition level of 96.0% which decreased to 76.0% of the COX-1 enzyme after 90 days of storage while aqueous extract showed 61.0% inhibition which decreased to 0% of the COX-1 enzyme after 90 days of storage [82]. Ndhkala *et al.* [2] investigated the anti-inflammatory effects of aqueous, dichloromethane, ethanol, and petroleum ether bulb extracts of *D. elata* using COX-1 and COX-2 inhibitory bioassays. The aqueous and ethanol extracts showed percentage inhibition of over 80.0% and 48.0%, respectively, for COX-1 while only the aqueous extract showed moderate inhibition of over 55.0% for COX-2 enzyme [2]. Ncube *et al.* [83] evaluated the anti-inflammatory activities of aqueous, dichloromethane, ethanol, and petroleum ether bulb and leaf extracts of *D. elata* collected in spring, summer, autumn, and winter seasons by assessing their ability to inhibit COX-1 and COX-2 enzymes. The dichloromethane and petroleum ether bulb and leaf extracts in all seasons except for winter when the leaves are not available showed moderate to high inhibition levels ranging from 58.0% to 94.1% of the COX-1 enzyme. A similar trend was observed for COX-2 enzyme with inhibition levels ranging from 52.8% to 91.2% [83]. These findings support the traditional use of *D. elata* as herbal medicine for back pain [39], body pains [2,3], inflammation [51], muscle pain [41], pain [43,46,51], and swelling [39].

#### Antioxidant activities

Matotoka and Masoko [70] evaluated antioxidant activities of acetone and hexane extracts of *D. elata* bulb using 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging assay. The hexane extracts exhibited antioxidant activities [70]. Matotoka and Masoko [43] evaluated antioxidant activities of an herbal mixture of *D. elata* bulb together with leaves of *M. angustifolia*, *S. viminale* and *Vahlia capensis*, *Kirkia wilmsii* (leaves, roots, and twigs), and *Hypoxis hemerocallidea* (corm) using the DPPH free radical scavenging assay and ferric reducing power with L-ascorbic acid as the positive control. The free radical scavenging activity showed that the herbal concoction exhibited moderate antioxidant activities. The ferric reducing power measuring the reduction of Fe<sup>3+</sup> to Fe<sup>2+</sup> revealed that the herbal concoction exhibited good reducing activity compared to L-ascorbic acid, the positive control [43]. The documented antioxidant activities of the bulb extracts of *D. elata* are probably due to flavonoids, gallotannins, phenolics, saponins, and tannins which have been isolated from the species [43,71,76].

#### Hemagglutinating activities

Gaidamashvili and Van Staden [84] evaluated hemagglutinating activities of aqueous bulb extracts of *D. elata* toward fresh and glutaraldehyde-treated rabbit erythrocytes using the hemagglutination and hapten inhibition assays. The extracts yielded hemagglutinating activity which was detected in the crude protein extracts at the minimal concentrations of 19.9 mg/ml. The was inhibited by 200 mM lactose along with major inhibition by D(+) trehalose, >DL arabinose, and D fructose [84]. The documented information on hemagglutinating activities and the identification of proteins from *D. elata* may be useful for future characterization of the species extracts in developing pharmaceutical products.

#### Cytotoxicity activities

Matotoka and Masoko [43] evaluated cytotoxicity activities of an herbal mixture of *D. elata* bulb together with leaves of *M. angustifolia*, *S. viminale* and *Vahlia capensis*, *Kirkia wilmsii* (leaves, roots, and twigs), and *Hypoxis hemerocallidea* (corm) using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide calorimetric assay with actinomycin D as the negative control. The cytotoxic concentration (CC<sub>50</sub>) values of all the concoctions were above the highest concentration used (1000 µg/mL) and Actinomycin D; the negative control exhibited CC<sub>50</sub> value of 0.6 µg/mL [43]. The documented cytotoxicity activities exhibited by *D. elata* extracts may be attributed to bufadienolides as these compounds are known to have cytotoxicity activities [64,65].

#### CONCLUSION

Based on information about *D. elata* that has been documented in this review, there appear to be research gaps on ethnopharmacological evaluation and clinical research on the species. No *in vivo* evaluations nor an assessment of target-organ toxicity have been carried out using the extracts from the species. Since *D. elata* is widely used in combination with other plant species in various herbal concoctions, there is a need for extensive research to evaluate synergistic effects of the different extracts and also to evaluate their ability to enhance the efficiency of the additive mixtures. Future research should also focus on aerial parts of the species to ensure full utilization of the possible medicinal potential of the species. Literature studies show that the major phytochemical compounds isolated from *D. elata* so far are mainly bufadienolides but very little attempt has been made to correlate the activities of these compounds with the ethnomedicinal uses of the species. Therefore, detailed phytochemical studies of *D. elata* and its pharmacological properties, especially the mechanism of action of its bioactive constituents to illustrate the correlation between its ethnomedicinal uses and pharmacological activities should be the focus of future research studies. Extensive *in vivo* experiments are required to validate the existing pharmacological activities. Since *D. elata* contain potentially toxic compounds, future studies should research on how potential toxic components of the species can be managed.

#### AUTHOR'S CONTRIBUTIONS

The author declares that this work was done by the author named in this article.

#### CONFLICTS OF INTEREST

The author declares that there are no conflicts of interest regarding the publication of this paper.

#### ACKNOWLEDGMENTS

The author would like to express his gratitude to the National Research Foundation (NRF), South Africa and Govan Mbeki Research and Development Centre (GMRDC), University of Fort Hare for financial support to conduct this study.

#### REFERENCES

1. Van Wyk BE, Oudtshoorn BV, Gericke N. Medicinal Plants of South Africa. Pretoria: Briza Publications; 2013.
2. Ndhkala AR, Finnie JF, Van Staden J. Plant composition, pharmacological properties and mutagenic evaluation of a commercial zulu herbal mixture: Imbiza ephuzwato. J Ethnopharmacol 2011;133:663-74.
3. Maroyi A. From traditional usage to pharmacological evidence: Systematic review of *Gunnera perpensa* L. Evidence Based Complement Altern Med 2016;2016:14.
4. Ndhkala AR, Van Staden J. Smokescreens and mirrors in safety and quality of herbal medicines: A case of commercialized herbal preparations. S Afr J Bot 2012;82:4-10.
5. Williams VL, Balkwill K, Witkowski ET. Unraveling the commercial market for medicinal plants and plant products on the Witwatersrand, South Africa. Econ Bot 2000;54:310-37.
6. Dold AP, Cocks ML. The trade in medicinal plants in the Eastern Cape Province, South Africa. S Afr J Sci 2002;98:589-97.

7. Loundou PM. Medicinal Plant Trade and Opportunities for Sustainable Management in the Cape Peninsula, South Africa. MSc Dissertation, Cape Town: University of Stellenbosch; 2008.
8. Cunningham AB. African Medicinal Plants: Setting Priorities at the Interface between Conservation and Primary Health Care. Paris: People and Plants Working Paper 1, UNESCO; 1993.
9. Williams VL. The Witwatersrand multi trade. Veld Flora 1996;82:12-4.
10. Williams VL, Balkwill K, Witkowski ET. A lexicon of plants traded in the Witwatersrand umuthi shops, South Africa. Bothalia 2001;31:71-98.
11. Ndawonde BG, Dlamini ET, Imenda SN. Indigenous knowledge, women and issues of sustainable development. WIT Trans Ecol Environ 2011; 167:45-54.
12. Moeng TE. An Investigation into the Trade of Medicinal Plants by Muthi Shops and Street Vendors in the Limpopo Province, South Africa. MSc Dissertation, Sovenga: University of Limpopo; 2010.
13. Petersen LM, Moll EJ, Collins R, Hockings MT. Development of compendium of local, wild-harvested species used in the informal economy trade, Cape Town, South Africa. Ecol Soc 2012;17:26.
14. Philander LE, Makunga NP, Esler KJ. The informal trade of medicinal plants by Rastafari bush doctors in the Western Cape of South Africa. Econ Bot 2014;68:303-15.
15. Ndawonde BG, Zobolo AM, Dlamini ET, Siebert SJ. A survey of plants sold by traders at Zululand muthi markets, with a view to selecting popular plant species for propagation in communal gardens. Afr J Range Forage Sci 2007;24:103-7.
16. Marshall NT. Searching for a Cure: Conservation of Medicinal Wildlife Resources in East and Southern Africa. Cambridge: TRAFFIC International; 1998.
17. Wiersum KF, Dold AP, Husselman M, Cocks ML. Cultivation of medicinal plants as a tool for biodiversity conservation and poverty alleviation in the Amatola region, South Africa. In: Bogers RJ, Craker LE, Lange D, editors. Medicinal and Aromatic Plants: Agricultural, Commercial, Ecological, Legal, Pharmacological and Social Aspects. Wageningen: Springer; 2006. p. 43-57.
18. Maroyi A, Mosina GK. Medicinal plants and traditional practices in peri-urban domestic gardens of the Limpopo province, South Africa. Indian J Tradit Knowl 2014;13:665-72.
19. Semanya SS, Potgieter MJ. Medicinal plants cultivated in bapedi traditional healers homegardens, Limpopo province, South Africa. Afr J Tradit Complement Altern Med 2014;11:126-32.
20. Mosina GK, Maroyi A, Potgieter MJ. Useful plants grown and maintained in domestic gardens of the Capricorn District, Limpopo province, South Africa. Stud Ethno Med 2015;9:43-58.
21. Azizi N, Amirouche R, Amirouche N. Cytotaxonomic diversity of some medicinal species of hyacinthaceae from Algeria. Pharmacogn Commun 2016;6:34-8.
22. Angiosperm Phylogeny Group (APG III). An update of the angiosperm phylogeny group. Classification for the orders and families of flowering plants: APG III. Bot J Linn Soc 2009;161:105-21.
23. Chase MW, Reveal JL, Fay MF. A subfamilial classification for the expanded asparagalean families, amaryllidaceae, asparagaceae and xanthorrhoeaceae. Bot J Linn Soc 2009;161:132-6.
24. Manning JC, Goldblatt P, Fay MF. A revised generic synopsis of hyacinthaceae in sub-Saharan Africa, based on molecular evidence, including new combinations and the new tribe pseudoprosperae. Edinburgh J Bot 2004;60:533-68.
25. Manning JC, Deacon J, Goldblatt P. A review of the schizobasis group of *Drimia* Jacq. (Hyacinthaceae: Urgineoideae), and the new species *D. sigmoidea* from Western Cape, South Africa. S Afr J Bot 2014;94: 263-9.
26. Jessop JP. Studies in the bulbous liliaceae in South Africa: 7. The taxonomy of *Drimia* and certain allied genera. J S Afr Bot 1977;43: 265-319.
27. Stedje B. A revision of the genus *Drimia* (Hyacinthaceae) in East Africa. Nordic J Bot 1987;7:655-66.
28. Stedje B. Hyacinthaceae. In: Polhill RM, editor. Flora of Tropical East Africa. Rotterdam: A. Balkema; 1996. p. 1-32.
29. Germishuizen G, Meyer NL. Plants of Southern Africa: An Annotated Checklist. Pretoria: Strelitzia 14, National Botanical Institute; 2003.
30. Figueiredo E, Smith GF. Plants of Angola. Pretoria: Strelitzia 22, South African National Biodiversity Institute; 2008.
31. Darbyshire I, Kordofani M, Farag I, Candiga R, Pickering H. The Plants of Sudan and South Sudan. London: Kew Publishing, Royal Botanic Gardens, Kew; 2015.
32. Cook FE. Economic Botany Data Collection Standard. London: Prepared for the International Working Group on Taxonomic Databases for Plant Sciences (TDWG), Kew Royal Botanic Gardens, Kew; 1995.
33. Macía MJ, Armesilla PJ, Cámara-Leret R, Paniagua-Zambrana N, Villalba S, Balslev H, et al. Palm uses in Northwestern South America: A quantitative review. Bot Rev 2011;77:462-570.
34. Gruca M, Cámara-Leret R, Macía MJ, Balslev H. New categories for traditional medicine in the economic botany data collection standard. J Ethnopharmacol 2014;155:1388-92.
35. World Health Organization (WHO). Atlas of African Health Statistics 2014. Geneva: Health Situation Analysis of the African Region, World Health Organization; 2014.
36. Gurib-Fakim A. Medicinal plants: Traditions of yesterday and drugs of tomorrow. Mol Aspects Med 2006;27:1-93.
37. Maroyi A. *Euclea crispa*: Review of its botany, ethnomedicinal uses and pharmacological properties. Asian J Pharm Clin Res 2018;11:5-9.
38. Maroyi A. *Dicoma anomala* Sond: A review of its botany, ethnomedicine, phytochemistry and pharmacology. Asian J Pharm Clin Res 2018;11:70-7.
39. Philander LA. An ethnobotany of Western Cape Rasta bush medicine. J Ethnopharmacol 2011;138:578-94.
40. Semanya SS, Potgieter MJ, Tshisikhawe MP. Use, conservation and present availability status of ethnomedicinal plants of Matebele-Village in the Limpopo Province, South Africa. Afr J Biotechnol 2013;12:2392-405.
41. Maema LP, Mahlo SM, Potgieter MJ. Ethnomedicinal uses of indigenous plant species in Mogalakwena municipality of Waterberg district, Limpopo province, South Africa. Int J Tradit Complement Med 2016;1: 28-44.
42. Masafu MM, Mbajorgu CA, Nematodzi LE, Kabine ES. A study of natural habitats and uses of medicinal plants in Thulamela and JS Moroka municipalities, South Africa. Indian J Tradit Knowl 2016;15: 363-9.
43. Matotoka MM, Masoko P. Phytochemical screening and pharmacological evaluation of herbal concoctions sold at Ga Maja Limpopo province. S Afr J Bot 2018;117:1-10.
44. Semanya SS, Wadesango N. Ethnobotanical survey of plants used by bapedi traditional healers to treat hypertension in the Polokwane municipality, Limpopo province, South Africa. Indilinga Afr J Indig Knowl Syst 2018;17:109-29.
45. Hutchings A, Scott AH, Lewis G, Cunningham AB. Zulu Medicinal Plants: An Inventory. Pietermaritzburg: University of Natal Press; 1996.
46. Long C. Swaziland's flora: siSwati Names and Uses. Swaziland National Trust Commission, Mbambane; 2005. Available from: <http://www.sntc.org.sz/index.asp>.
47. Van Wyk BE, Gericke N. Peoples Plants: A Guide to Useful Plants of Southern Africa. Pretoria: Briza Publications; 2007.
48. Semanya SS, Maroyi A. Ethnobotanical survey of plants used by bapedi traditional healers to treat tuberculosis and its opportunistic infections in the Limpopo Province, South Africa. S Afr J Bot 2018. Doi: 10.1016/j.sajb.2018.10.010.
49. Hutchings A. Plants Used for Some Stress-related Ailments in Traditional Zulu, Xhosa and Sotho Medicine. MSc Dissertation. Pietermaritzburg: University of Natal; 1992.
50. Madisha JK. Antimycobacterial Activities of Selected Plants Used in the Management of Tuberculosis in Sekhukhune (Limpopo Province), South Africa. MSc Dissertation. Bloemfontein: University of the Free State; 2017.
51. Hulley IM, Van Wyk BE. Quantitative medicinal ethnobotany of Kannaland (Western Little Karoo, South Africa): non-homogeneity amongst villages. S Afr J Bot 2018. Doi: 10.1016/j.sajb.2018.03.014.
52. Erasmus LJ, Potgieter MJ, Semanya SS, Lennox SJ. Phytomedicine versus gonorrhoea: The bapedi experience. Afr J Tradit Complement Altern Med 2012;9:591-8.
53. Semanya SS, Potgieter MJ, Erasmus LJ. Indigenous plant species used by bapedi healers to treat sexually transmitted infections: Their distribution, harvesting, conservation and threats. S Afr J Bot 2013;87: 66-75.
54. Semanya SS, Potgieter MJ, Erasmus LJ. Ethnobotanical survey of medicinal plants used by bapedi traditional healers to manage HIV/AIDS in the Limpopo province, South Africa. J Med Plants Res 2013; 7:434-41.
55. Semanya SS, Potgieter MJ, Erasmus LJC. Bapedi phytomedicine and their use in the treatment of sexually transmitted infections in Limpopo province, South Africa. Afr J Pharm Pharmacol 2013;7:250-62.
56. Semanya SS, Potgieter MJ. Ethnobotanical survey of medicinal plants used by bapedi traditional healers to treat erectile dysfunction in the Limpopo Province, South Africa. J Med Plants Res 2013;7:349-57.
57. Semanya SS, Maroyi A, Potgieter MJ, Erasmus LJ. Herbal medicines used by bapedi traditional healers to treat reproductive ailments in the Limpopo

- Province, South Africa. Afr J Tradit Complement Altern Med 2013; 10:331-9.
58. Pujol J. Natur Africa: The Herbalist Handbook, African Flora, Medicinal Plants. Durban: Jean Pujol Natural Healers Foundation; 1990.
  59. Kellerman TS, Coetzer JA, Naudé TW. Plant Poisonings and Mycotoxicoses of Livestock in Southern Africa. Cape Town: Oxford University Press; 1988.
  60. Kellerman TS, Naudé TW, Fourie N. The distribution, diagnosis and estimated economic impact of plant poisonings and mycotoxicoses in South Africa. Onderstepoort J Vet Res 1996;63:65-90.
  61. Van Wyk BE, Van Heerden F, Van Oudtshoorn B. Poisonous Plants of South Africa. Pretoria: Briza Publications; 2005.
  62. Kupchan SM, Hemingway RJ, Hemingway JC. The isolation and characterization of hellebrigenin 3-acetate and hellebrigenin 3,5-diacetate, bufadienolide tumor inhibitors from *Bersama abyssinica*. Tetrahedron Lett 1968;2:149-52.
  63. Kupchan SM, Ognyanov I, Moniot JL. Tumor inhibitors. 64. Isolation and structural elucidation of novel bufadienolides, the cytotoxic principles of *Bersama abyssinica*. Bioorg Chem 1971;1:13-31.
  64. Yamagishi T, Yan XZ, Wu RY, McPhail DR, McPhail AT, Lee KH, et al. Structure and stereochemistry of bryophyllin-A, a novel potent cytotoxic bufadienolide orthoacetate from *Bryophyllum pinnatum*. Chem Pharm Bull (Tokyo) 1988;36:1615-7.
  65. Yamagishi T, Haruna M, Yan XZ, Chang JJ, Lee KH. Antitumor agents, 110. bryophyllin B, a novel potent cytotoxic bufadienolide from *Bryophyllum pinnatum*. J Nat Prod 1989;52:1071-9.
  66. Supratman U, Fujita T, Akiyama K, Hayashi H, Murakami A, Sakai H, et al. Anti-tumorpromoting activity of bufadienolides from *Kalanchoe pinnata* and *K. daigremontiana* Xbutiflora. Biosci Biotechnol Biochem 2001;65:947-9.
  67. Moodley N, Crouch NR, Mulholland DA. Bufadienolides from *Drimia macrocentra* and *Urginea riparia* (Hyacinthaceae: Urgineoideae). Phytochemistry 2007;68:2415-9.
  68. Dai Y, Harinantenaina L, Brodie PJ, Goetz M, Shen YY, Dyke KT, et al. Antiproliferative flavonoids and bufatrienolides from *Urginea depressa*. J Nat Prod 2013;76:865-72.
  69. Koorbanally NA, Koorbanally C, Harilal A, Mulholland DA, Crouch NR. Bufadienolides from *Drimia robusta* and *Urginea epigea* (Hyacinthaceae). Phytochemistry 2004;65:3069-73.
  70. Matotoka MM, Masoko P. Evaluation of herbal concoctions sold at Ga Maja (Limpopo Province) in South Africa and *in vitro* pharmacological evaluation of plants used to manufacture the concoctions. J Evidence Based Complement Altern Med 2017;22:805-15.
  71. Okem A, Southway C, Stirk WA, Street RA, Finnie JF, Van Staden J. Heavy metal contamination in South African medicinal plants: A cause for concern. S Afr J Bot 2014;93:125-30.
  72. Luyt RP, Jäger AK, Van Staden J. The rational usage of *Drimia robusta* Bak. In traditional medicine. S Afr J Bot 1999;65:291-4.
  73. Luyt RP, Jäger AK, Van Staden J. Bufadienolides in *in vitro* derived *Drimia robusta* plants. S Afr J Bot 1999;65:443-5.
  74. Krenn L, Stapf V, Kopp B. Bufadienolides from *Drimia robusta* BAK. Sci Pharm 2000;68:421-7.
  75. Pohl T, Koorbanally C, Crouch NR, Mulholland DA. Bufadienolides from *Drimia robusta* and *Urginea altissima* (Hyacinthaceae). Phytochemistry 2001;58:557-61.
  76. Ncube B, Finnie JF, Van Staden J. Seasonal variation in antimicrobial and phytochemical properties of frequently used medicinal bulbous plants from South Africa. S Afr J Bot 2011;77:387-96.
  77. Baskaran P, Singh S, Van Staden J. *In vitro* propagation, proscillaridin A production and antibacterial activity in *Drimia robusta*. Plant Cell Tissue Organ Cult 2013;114:259-67.
  78. Okem A, Southway C, Stirk WA, Street RA, Finnie JF, Van Staden J. Effect of cadmium and aluminum on growth, metabolite content and biological activity in *Drimia elata* (Jacq.) Hyacinthaceae. S Afr J Bot 2015;98:142-7.
  79. Baskaran P, Kumari A, Van Staden J. Analysis of the effect of plant growth regulators and organic elicitors on antibacterial activity of *Eucomis autumnalis* and *Drimia robusta* *ex vitro*-grown biomass. Plant Growth Regul 2018;85:143-51.
  80. Kandari LS. Effect of vermicompostleachate on biomass and antibacterial properties of five bulbous medicinal plants: A case study from Kwa Zulu Natal, South Africa. Iran J Sci Technol Trans A Sci 2018;42:1049-56.
  81. Fouche G, Gragg GM, Pillay P, Kolesnikova N, Maharaj VJ, Senabe J. *In vitro* anticancer screening of South African plants. J Ethnopharmacol 2008;119:455-61.
  82. Stafford GI, Jäger AK, Van Staden J. Effect of storage on the chemical composition and biological activity of several popular South African medicinal plants. J Ethnopharmacol 2005;97:107-15.
  83. Ncube B, Finnie JF, Van Staden J. Seasonal variation in the cyclooxygenase inhibitory activities of four South African medicinal bulbs. S Afr J Bot 2012;78:246-51.
  84. Gaidamashvili M, Van Staden J. Lectin-like proteins from South African plants used in traditional medicine. S Afr J Bot 2002;68:36-40.