

NOVEL APPROACHES FOR DEVELOPMENT OF ORAL CONTROLLED RELEASE COMPOSITIONS OF GALANTAMINE HYDROBROMIDE AND PAROXETINE HYDROCHLORIDE HEMIHYDRATE: A REVIEW

UMESH NANDKUMAR KHATAVKAR^{1,2,3}, K. JAYARAM KUMAR², SHAMKANT LAXMAN SHIMPI¹

¹Formulation Research Division, Aurobindo Pharma Limited Research Centre, 313, Bachupally, Quthubullapur, Hyderabad 500072, India,

²Department of Pharmaceutical Sciences, Birla Institute of Technology, Mesra Ranchi, India, ³To whom correspondence should be addressed

Email: khatavkarun@yahoo.com

Received: 16 Feb 2016, Revised and Accepted: 20 Jun 2016

ABSTRACT

The objective of this review is to study different novel approaches for achieving controlled release for oral administration. There is need of developing cost-effective generic products which will be comparable to the established innovator products with respect to *in vivo* performance. The innovator products being developed based on exhaustive research are developed utilizing novel platform technologies, have been protected with patents. These platform technologies require specialized manufacturing equipment's and that additionally imparts overall cost to the drug product. This review also contains review on existing technologies utilized for controlling the drug release of actives and also focuses on authors work on the development of cost-effective novel approaches for developing controlled release of some selected central nervous system acting drugs viz galantamine hydrobromide (GAH) and paroxetine hydrochloride hemihydrate (PHH). The existing approved reference products of selected molecules are available based on extended release multi-particulate delivery system and Geomatrix based platform technology for GAH and PHH respectively. This review also explains authors work in developing different controlled release approaches for achieving similar *in vivo* performance comparable to the reference product. The use of high viscosity grade of hydroxypropyl cellulose (HPC) as a release controlling matrix former in order to control the release of GAH by direct compression into mini tablets offers a feasible dosage form which further can be filled into capsules. Hydroxypropylmethylcellulose (HPMC) based matrix tablets which were further coated using methacrylic acid copolymer were found to be a suitable method to formulate single layer controlled release PHH.

Keywords: Paroxetine hydrochloride hemihydrates, Galantamine hydrobromide, Aquacoat, Hydroxypropyl methylcellulose, Geomatrix technology, *In vivo*

© 2016 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>)

INTRODUCTION

Development of pharmaceutical product formulation in a timely manner and ensuring quality is a complex process that requires a systemic, science-based approach. Information from various categories, including the property of drug substance and excipients, the interaction between material, unit operation, and equipment is gathered. Knowledge in different forms, including heuristics, decision trees, correlation, and first principle model is applied. Decisions regarding processing routes choice of excipients and equipment sizing are made based on this information and knowledge. Controlled drug delivery technology has improved over the last few decades. The controlled release system provides a constant supply of the active ingredient *in vivo*, usually at a zero-order rate, by constantly releasing, for a certain period of time, an amount of the drug approximately similar to the amount that is eliminated by the body. Literature review has been carried out from 1907 to till date through Chemical abstract service, Sci Finder, Google scholar, Pub Med, a library search and manual search of traditional as well as contemporary books to evaluate physicochemical, pharmacokinetic, properties of Galantamine hydrobromide and Paroxetine hydrochloride hemihydrate, their reported formulation and pharmacokinetic evaluation properties.

Different approaches for controlled release formulations

Conte *et al.* (1993) disclosed multilayer tablet systems for controlling the drug release. They disclosed novel approach as against conventional coating approach to achieve constant drug release. The novel design where the matrix core containing active substance is restricted from contact with dissolution medium which impact in the rate of hydration & swelling of polymeric matrix and the release of active is controlled in a constant manner. By increasing the extent of a barrier to the active surface, systems performs the drug towards constant rate [1].

Conte *et al.* (1996) further studied and evaluated Geomatrix platform technology for drugs having different solubility. They

studied a different type of barrier layers viz swellable and erodible. Authors studied the effect of different barrier property polymer systems with trapidil, ketoprofen and nifedipine. The study revealed that swellable polymeric barrier suits for active substances having high solubility while erodible polymeric barrier suits for sparingly soluble actives [2].

Rodriguez and Vila-Jato (1998) developed multi particulate system containing hydrophobic core which is further coated with pH dependent eudragit polymer in order to target the release of active in the colon. The system consisted of drug loaded cellulose acetate butyrate microspheres which further coated with Eudragit. The dissolution study revealed pH dependent release controlled by eudragit while the further release was controlled by cellulose acetate butyrate [3].

Khan and Jiabi (1998) evaluated sustained release matrix tablets of ibuprofen using carbopol as a release controlling polymer. The formulations were prepared by direct compression method using different concentrations of carbopol. The drug release rate was found to be dependent on carbopol content in the formulation. As the content is increased the rate of release was found to be retarded. Effect of other excipients was also studied. The data revealed that formulation containing lactose shown to be slower and linear drug release than microcrystalline cellulose and starch [4].

Eftentakis *et al.* (2000) evaluated the difference in release mechanism using carbopol and sodium alginate as release-controlling excipients. Single unit tablets and capsules and multi-unit mini tablets filled in capsules were prepared and the dissolution behavior was studied. The carbopol containing formulations were shown more swelling and less erosion as against sodium alginate [5].

Varma and Garg (2001) reported different therapeutic objectives that can met using different Geomatrix technologies i) Constant or near zero order drug release ii) binary drug release for a combination of drugs from same tablet iii) positioned delivery of active at a predetermined location in the body iv) delayed release [6].

Krishnaiah *et al.* (2002) evaluated three layer matrix controlled release tablet formulation of trimetazidine dihydrochloride using guar gum. The matrix granules were prepared by wet granulation. The impact of different concentration of guar gum in active layer and support layers was studied. The formulation showed dissolution stability when studied for accelerated storage condition [7].

Tiwari *et al.* (2003) evaluated hydrophilic and hydrophobic matrix system for tramadol hydrochloride. Hydrophilic matrix tablets were prepared by wet granulation using HPMC while hydrophobic matrix tablets were prepared using melt granulation. Hydrophobic matrix system was found to control the release rate effectively [8].

Rao, Engh and Qui (2003) studied the effect of pH modifiers on the release rate of acidic drug divalproex sodium. Authors studied the effect of eudragit E 100 and fujicalin. The formulation containing eudragit E 100 was found to show pH independent drug release. This was attributed to elevation of microenvironmental pH [9].

Royce *et al.* (2004) studied *in vitro in vivo* correlation between three different controlled release systems. The model drug was formulated in osmotic, hydrophilic matrix and reservoir technique. Although all systems showed similar dissolution profile, the *in vivo* outcome was different. Out of three controlled release systems osmotic system shown good co relation of dissolution and *in vivo* performance [10].

Korhonen *et al.* (2004) studied *in vivo* performance of three different release rate systems in humans. The different release rate formulations of diltiazem were prepared using starch acetate. Good co relation was observed between *in vitro* drug release rate/extent and *in vivo* parameters such as C_{max} and AUC. The slowest release formulation was observed to be equivalent with reference formulation [11].

Kim C (2005) studied dissolution of donut shaped three layered tablets. The matrix-based model drug core was prepared using enteric polymer HPMC acetate succinate. The top and bottom surface of core tablets were made of hydrophobic polymer ethyl cellulose. The system is reported to be useful in achieving near zero order release [12].

Jamzad and Fassih (2006) studied *in vitro* drug release of glipizide by preparing two different hydrophilic matrix systems using hydroxypropyl methylcellulose and polyethylene oxide. Matrix polymer hydration, erosion were evaluated during dissolution study. The dissolution data was compared with reference product Glucotrol XL based on osmotic drug delivery. The matrix based on HPMC shown near zero order drug release when compared to reference formulation [13].

Lopes *et al.* (2006) studied sustained release of ibuprofen from mini tablets. The controlled release of ibuprofen was achieved by preparing minitables using HPMC and ethyl cellulose. The geometry of mini tablets plays an important role in drug release rate. Because of the small size of the mini tablets initial release of drug from the surface of tablets was found to be more as compared to large tablet [14].

Siepmann *et al.* (2008) reviewed various polymeric blends that can be used to achieve controlled release of actives. Different polymeric blends can be used for a) desired drug release b) drug release mechanism c) achieving site specific release of active in the gastrointestinal tract. However, authors reported that these polymeric blend systems are more complex and chemical as well as physical compatibility between blend components need to be checked [15].

Ishida *et al.* (2008) studied sustained release of pseudoephedrine hydrochloride by preparing an immediate release and sustained release minitables. The drug release for sustained release component was achieved by coating using a combination of hydrophilic and hydrophobic polymers [16].

Dey Majumdar and Rao (2008) reported that because of a smooth surface, uniform surface area and high strength, minitables provide an advantage over granules or pellets wherein less amount of release controlling polymer is required to attain sustained release of active [17].

Baloglu and Senyigit (2010) studied multi-layered matrix tablet formulations of metoprolol tartrate. Different swellable hydrophilic polymers were used in combination for controlling the drug release. The dissolution data was mapped with the target release profile determined from reported pharmacokinetic data. The three layer matrix tablet system using carrageenan as a release controlling polymer was found to have comparable drug release profile with target drug release profile [18].

Abuelwafa and Basalious (2010) studied different approaches for controlling drug release of venlafaxine hydrochloride. Hydrophilic matrix based system, wax based matrix and three-layered tablets containing hydrophobic core sandwiched between hydrophilic matrix layers. The authors optimized the formulation performance using factorial design. Further, the optimized formulation was studied for *in vivo* bioequivalence study using reference capsule product as a reference. The test product was shown to be bioequivalent based on AUC parameter [19].

Rao *et al.* (2011) studied the use of mini tablets approach for achieving sustained release of montelukast sodium. Approximately 30 % of the drug release was achieved within 1 hour by preparing immediate release minitabled portion while remaining 70% of the drug release was achieved up to 24 hours by preparing differentially coated minitables [20].

Mamani *et al.* (2012) studied the effect of the different ratio of hydrophilic polymer HPMC and anhydrous dibasic calcium phosphate on theophylline release rate. Both the components in the matrix tablets have shown the impact on drug release across different pH of dissolution medium [21].

Literature on galantamine, paroxetine and author's research contribution

A. Galantamine hydrobromide

Mashkovsky and Kruglikova-Lvova (1951) isolated and studied acetylcholinesterase (AChE)-inhibiting properties of galantamine [22]. Proskumina and Yakovleva (1952) reported the isolation of an alkaloid; GAH from bulbs of the caucasian snowdrops *Galanthus woronowii* [23]. Davis (1987) disclosed method of treating Alzheimer's disease using parenteral administration of GAH [24]. GAH is a reversible, competitive acetylcholinesterase inhibitor used for the treatment of mild to moderate dementia of the Alzheimer's type. The efficacy of GAH has been reported in patients with severe Alzheimer's disease. GAH is a tertiary alkaloid, belonging to the phenanthrene chemical class [25]. GAH as per biopharmaceutical classification system would be classified as class 1 drug substance. GAH extended-release capsule dosage form has been approved by United States Food and Drug Administration (USFDA) in April 2005 and is commercially available under the trade name Razadyne ER® in the United State and available in 8 mg, 16 mg and 24 mg strengths. It is benzazepine derivative having ionization constant of 8.2. Its melting point is reported to be 246-247 °C. There are no polymorphic forms reported in the literature. [26].

The Razadyne ER® as disclosed in US Patent 7,160,559 comprises the three stage coating process to achieve the desired release profile. The product is based on reservoir controlled release multi particulate delivery system. The first stage comprises coating of GAH and water soluble film forming a polymer on inert spheres. The second stage comprises a further coating of a release rate controlling polymer membrane consisting of a combination of water soluble and water insoluble polymers. To achieve the desired release profile the third top coat is given consisting of water soluble polymer and GAH wherein the formulation is capable of releasing 20 to 40% of the total amount of GAH in 1 hour, and more than 80% of the total amount of GAH in 10 h [27]. The reference product (Razadyne ER® capsules) is based on multi particulate capsule dosage formulation. The manufacturing process involves many complicated steps and use of sophisticated equipment including fluid bed processor.

Kays *et al.* (2007) reported optimization of an intra-nasal GAH formulation using an *in vitro* tissue model, to correlate those results to *in vivo* bioavailability, and to compare emetic response to oral dosing. GAH permeation was enhanced without increasing

cytotoxicity. Pharmacokinetic testing in rats confirmed the improved drug bioavailability and demonstrated an *in vitro*-*in vivo* correlation. Compared to oral dosing, intranasal GAH resulted in a dramatically lowered incidence of GI-related side effects, e. g., retching and emesis [28].

Caro et al. (2009) disclosed the tablets designed for GAH administration on the buccal mucosa, were prepared by direct compression of drug loaded eudragit® RS 100 matrices. When the tablets were coated with a lipophilic material, GAH is slowly discharged from buccal tablets, following the Higuchian kinetic [29].

Controlled drug delivery of GAH which is based on reservoir multi particulate pellet technology similar to reference formulation involving one stage coating has been reported. The researchers studied dissolution behavior of the extended release multi particulate drug delivery system by preparing pellets of GAH using extrusion/spheronization. Eudragit RS 30D & RL 30 D were used as release-retarding polymers. The test formulations were reported to follow first order release rate kinetics [30].

Sharma and Biswal (2013) are evaluated bilayer tablet approach for developing sustained release of GAH. Immediate release as a loading dose and sustained release in the second layer were evaluated using HPMC as release retarding polymer. The sustained release was achieved over a period of 24 h [31].

Woo et al. (2015) evaluated transdermal patch system for delivery of GAH using carbopol polymer. The trial formulations were optimized using response surface methodology. The delivery system was reported to deliver cumulative release at 8 h [32].

Rao et al. (2015) studied different polymeric blends of GAH using konda gogu gum, sodium alginate and crosslinking agents. The sustained drug release was observed up to 12 h [33].

The authors have also developed controlled release compositions of GAH using a matrix based monolithic and reservoir based system [34, 35].

In one of the approaches based on reservoir technique, desired release profile of GAH was targeted by employing preparation of inert core mini tablets and coating of 70 % of GAH over the inert core, further controlling the release by coating composition consisting hydrophobic and hydrophilic components followed by a final coating of 30 % immediate dose of GAH. The controlled release mini tablets further filled into empty hard gelatin capsules.

The formulation G1 was manufactured by sifting lactose monohydrate and dibasic calcium phosphate dihydrate through 425 µm sieve. The sifted powder blend was then mixed for 15 min in an octagonal blender. Magnesium stearate was then sifted through 250 µm sieve and blended for 5 min in an octagonal blender. For formulation G2 the powdered blend of lactose monohydrate and dibasic calcium phosphate dihydrate was granulated by using 0.1 N hydrochloric acid in rapid mixer granulator, dried at 50 °C using the rapid dryer and sifted through 425 µm sieve. Magnesium stearate was then sifted through 250 µm sieve and blended for 5 min in an octagonal blender. Formulation G3 was manufactured by sifting mannitol and dibasic calcium phosphate dihydrate through 425 µm sieve. The sifted powder blend was then mixed for 15 min in an octagonal blender and further granulated by a solution of povidone in rapid mixer granulator, dried at 50 °C using the rapid dryer and sifted through 425 µm sieve. Magnesium stearate was then sifted through 250 µm sieve and blended for 5 min in an octagonal blender. The lubricated blends of different formulations were further used for compression into mini tablets using 10 stations single rotary compression machine. Further, these mini tablets were coated using 70 % of the dose of GAH in 10-inch gans coater.

The release controlling polymer solution of a different ratio of ethyl cellulose: hypromellose: diethyl phthalate was prepared by dissolving into ethanol (For G1-62:21:17, for G2 and G3-58:21:21). Finally, the remaining 30 % of the dose of GAH was loaded by coating over controlled release reservoir mini tablets. Four mini tablets were filled in capsule shells for 8 mg strength. The round standard concave punch & die of 4.5 mm diameter was used.

The drug release profile of GAH from the mini tablets filled in capsules and the reference formulation (Razadyne ER® capsules) in pH 6.5 phosphate buffer using paddle apparatus are shown in fig. 1. The drug release showed fast dissolution during the first hour because of 30% of GAH in immediate release portion. The core mini tablets prepared using 0.1N hydrochloric acid (G2) showed pH independent dissolution as against the core mini tablets devoid of 0.1N hydrochloric acid (G1 and G3). Further, the amount of release controlling polymer required was slightly more in the case of mini tablets containing increased amount of hydrophilic component (G3 formulation containing 76% of mannitol as filler) as compared to mini tablets containing less amount of hydrophilic component (G2 formulation containing 39.6% of lactose monohydrate as filler).

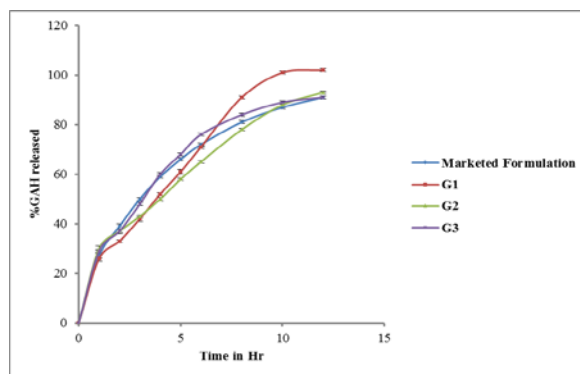


Fig. 1: % GAH released from mini reservoir tablets filled in capsules (G1-G3)®

The drug release of GAH from the formulations under investigations and the reference formulation was further studied in two different dissolution medias viz 0.1 N hydrochloric acid and pH 4.5 Acetate buffer in order to study the impact of pH of dissolution medium on the rate of drug release. The data is shown in fig. 2.

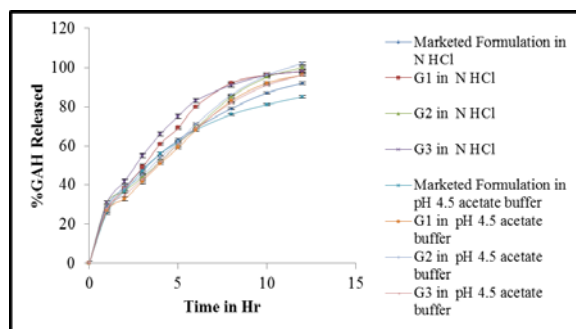


Fig. 2: % GAH released from formulations (G1-G3) based on reservoir approach and reference formulation in 0.1N HCl and pH 4.5 acetate buffer#

The reference formulation showed pH-independent release of the drug while the formulation G1 and G3 shown slightly increased the rate of drug release in pH 1.2. The formulation G2 with acidified core is shown comparable drug release profile to reference formulation in all Medias studied.

In another approach of developing controlled release of GAH, based on hydrophilic monolithic matrix mini tablets approach to control and achieve the desired release profile of GAH as comparable with Razadyne ER® capsules. The simple, direct compression technique was employed to formulate the mini tablets, which can be filled in capsules. The mini tablets were formulated with a hydrophilic polymer, HPC and GAH.

The formulations G1 and G2 with (46.66%, and 61% of HPC) were manufactured by sifting GAH, lactose monohydrate, HPC, and colloidal silicon dioxide through 425 μm sieve. The sifted powder blend was then mixed for 15 min in an octagonal blender. Magnesium stearate was then sifted through 250 μm sieve and mixed with previously blended drug excipient blend for 5 min in an octagonal blender. For formulations G3, G4 and G5 (with 76.49 %, 78.56 % and 83.31%) additionally talc was used as a lubricant. The lubricated blends of different formulations were further used for compression into mini tablets using 10 stations single rotary compression machine. Further, these mini tablets were filled into empty hard gelatin capsule shells of size 1. One mini tablet was filled in capsule shells for 8 mg strength. The round standard concave punch and die of 5.2 mm diameter was used.

The dissolution profile of GAH from the mini tablets filled in capsules and the reference formulation (Razadyne ER® capsules) in pH 6.5 phosphate buffer using paddle I apparatus are shown in fig. 3. The release patterns of reference formulations showed fast dissolution and burst effect during the first hour. This release pattern is due to immediate release top coat on the pellets as disclosed in US Patent 7,160,559. The release of reference formulation after the first hour was further found to be slowed down due to release rate controlling polymer membrane coating of a combination of water soluble and water insoluble polymers. The similar release pattern was observed for the developed test formulation without having immediate release portion in the formulation.

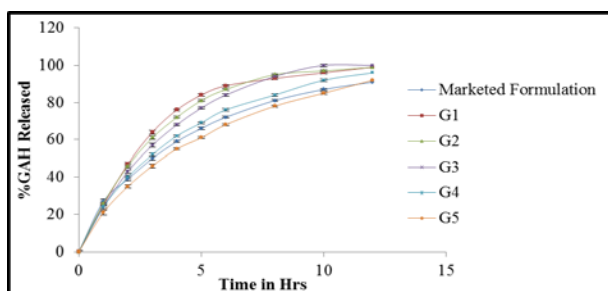


Fig. 3: Dissolution profile of test (G1-G5) and reference formulations[^]

The drug release of GAH from the formulation G5 and the reference formulation was studied in two different dissolution medias viz 0.1 N hydrochloric acid and pH 4.5 acetate buffer in order to study the impact of pH of dissolution medium on the rate of drug release. The data is shown in fig. 4. The reference formulation shown pH-independent release of the drug while the formulation G5 shown slightly increased the rate of drug release in pH 1.2. The reason could be due to the property of HPC which is prone for hydrolysis in acidic pH.

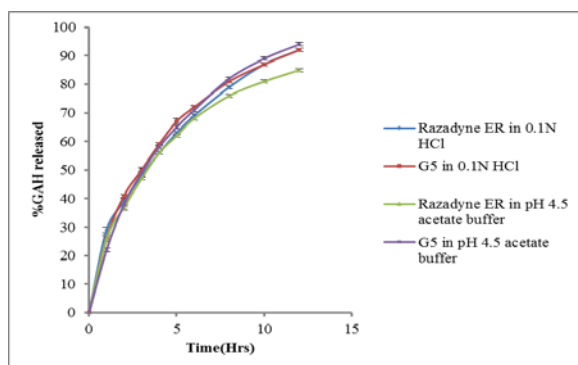


Fig. 4: % GAH released from matrix mini tablets (G5) filled in capsules in 0.1N HCl and pH 4.5 acetate buffers^{^^}

Based on the two approaches for controlling the release of GAH, it was observed that mini tablets based on a hydrophilic matrix containing HPC as a release controlling polymer was easy from manufacturability perspective & also cost effective. Authors evaluated the test formulation based on hydrophilic matrix based mini tablets for *in vivo* bioequivalence study & concluded the bioequivalence of test formulation with reference formulation.

B. Paroxetine hydrochloride hemihydrate

PHH is a serotonin re-uptake inhibitor useful for the treatment of psychiatric problems including depression, Parkinson's disease, anxiety disorders, obsessive-compulsive disorders, panic disorder and post-traumatic stress disorder. PHH belong to phenylpiperidine chemical class [36]. It is reported to exhibit different polymorphic forms. PHH controlled release tablet dosage form has been approved by United States Food and Drug Administration (USFDA) in Feb 1999 and is commercially available under the trade name PAXIL CR® tablets and available in 12.5 mg, 25 mg and 37.5 mg strengths. The reference product PAXIL CR® as disclosed in US Patent 4839177 and U. S. 5, 422, 123 is based on Geomatrix™ platform technology comprising bilayer tablet (Drug containing matrix layer & hydrophobic placebo matrix layer) further coated with enteric polymer release of PHH predominantly in the small intestine (zero order kinetics). The manufacturing process involves many complicated steps and use of sophisticated equipment including bilayer tablet compression machine.

Raghupathi *et al.* (2007) evaluated single layer delayed and controlled release tablets of PHH as against bilayer tablets of the reference product. The *in vivo* study resulted in an enhanced bioavailability of the test formulation suggesting for reducing the dose of the formulation [37].

Jin *et al.* (2008) studied the effects of the formulation variables-POLYOX molecular weight, the ratio of POLYOX/Avicel PH102 and the amount of POLYOX and Avicel PH102, hardness, HPMCP amount, eudragit L100 amount, and citric acid amount on the PHH release from POLYOX matrix tablet using the Plackett-Burman screening design. POLYOX molecular weight had significant influence on the drug release mechanism [38].

Vaidya *et al.* (2013) evaluated controlled release of paroxetine using high viscosity HPMC polymers and compritol ATO. The formulation with desired release profile was shown stability in dissolution for one month [39].

The authors have also studied different approaches of controlled release of PHH. In one approach the preparation of single layer controlled release containing hydrophilic matrix core was attempted wherein the release from matrix core tablets was further delayed using methacrylic acid copolymer dispersion. In this approach, 85% of the dose was included in core matrix whereas remaining 15% of the dose was included as an immediate release as an outer coating layer over delayed release coating. In another approach, hydrophobic matrix core containing methacrylic acid copolymer was prepared by wet granulation in order to delay and control the drug release. In this approach, 100% of the dose was included in the core matrix. In yet another approach the drug substance PHH was granulated with methacrylic acid copolymer dispersion and further hydrophilic matrix tablets were prepared

The formulations P1-P8 were prepared by wet granulating 85 % of the dose with hydrophilic polymer & other excipients. The compressed matrix core tablets were further coated with an enteric polymer. The enteric coated tablets were further coated with 15 % of the dose as a top coat. The formulations P4 and P5 were further coated with ethylcellulose aqueous dispersion. The formulations P9-P12 were prepared by granulating 100 % of the dose, hydrophobic matrix components along with methacrylic acid copolymer by wet granulation and compressed into tablets. The formulations P13-P16 were prepared by using PHH granulated with 4% (P13 and P14) and 8% (P15 and P16) methacrylic acid copolymer dispersion. The granulated PHH was further used for preparing hydrophilic matrix core tablets [40].

The dissolution profile of PHH from controlled release matrix tablets and the reference formulation (PAXIL CR® tablets) in USFDA recommended dissolution methodology (Paddle, 150 RPM); paddle apparatus at 75 RPM and basket apparatus at 100 RPM are shown in fig. 5.1, 5.2 and 5.3. The reference formulation showed no drug release in 0.1 N hydrochloric acid for 2 h in all dissolution methodologies because of the presence of methacrylic acid copolymer coating which has been reported to be solubilized at a pH of 5.5 and above.

Whereas the formulations (P1-P8) showed the release of up to 15% in 0.1 N hydrochloric acid for 2 h in all dissolution methodologies which indicated the enteric coating over 85% of the dose prevented further drug release from the core matrix. The formulation P9 prepared with granulating the hydrophilic core matrix containing methacrylic acid copolymer showed 19% drug release in 0.1 N hydrochloric acid. The release in acidic medium was controlled because of the methacrylic acid copolymer in the matrix. The formulations P10 and P12 containing varied concentrations of methacrylic acid copolymer showed slower drug release profile. It could be because of the hydrophobic effect of glyceryl behenate in the formulation. The drug release of about 6-10% was observed in acidic medium. The formulation P11 was similar to P10 with additional hydrophilic polymer added at extragranular stage significantly enhanced the drug release as compared to formulations P10 and P12. The formulation was unable to maintain the matrix integrity, and initial burst release of 66% in acidic medium was observed which further released 93% immediately in 1st hour in buffer medium.

The formulations containing methacrylic acid copolymer granulated PHH within hydrophilic matrix (P13-P16) also showed controlled drug release. The formulations containing 100 % of the dose granulated with the methacrylic acid copolymer (P13 and P16) showed slightly slower release profile as compared to the formulations P14 and P16 containing 50 % of the dose granulated with the methacrylic acid copolymer. There was no significant difference observed within P13-P16 for PHH release in acidic medium and all formulations showed PHH release of about 20-24% in acidic medium. Although there was no significant difference in the rate and extent of PHH drug release amongst the formulations P13-P16, surprisingly the formulations containing 4 % granulated PHH (P13 and P14) showed slightly slower drug release as compared to formulations containing 8% granulated PHH (P15 and P16).

The release of reference and test formulations in pH 6.8 phosphate buffers in paddle at 150 RPM was further found to be controlled due to hydration of polymer HPMC. The increase in the concentration of polymer HPMC from 8.86 % to 14.29% based on the weight of core matrix composition (P1, P6, P7 and P8) resulted in a decrease in the rate of drug release (Figure. 5). This is attributed to gel layer formation with a longer diffusion path as the content of HPMC was increased.

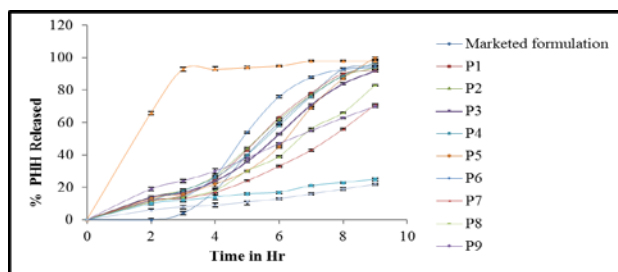


Fig. 5: Comparative PHH release profile between test formulations (P1-P9) and reference formulation (Paddle 150RPM)^{ss}

Based on the data from different approaches, the use of high viscosity grade of HPMC as a release controlling matrix former in order to control the release of PHH by wet granulation into single layer matrix tablet with further delayed release coat using methacrylic acid copolymer dispersion offers a feasible dosage form.

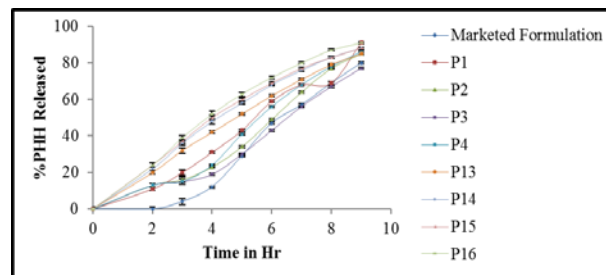


Fig. 6: Comparative PHH release profile between test formulations (P1-P4, P13-P16) and reference formulations (Basket 100 RPM)^{ss}

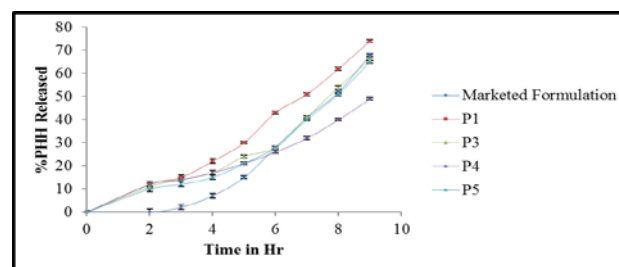


Fig. 7: Comparative PHH release profile between test formulations (P1, P3, P4, P5) and reference formulations (Paddle 75 RPM)^{ss}

The matrix tablets prepared by granulating with methacrylic acid copolymer also showed promising strategy and needs to be further evaluated and optimized in order to achieve comparable dissolution with reference formulation. The development approach involving granulated PHH in the hydrophobic matrix have shown to be less promising.

CONCLUSION

Reference formulation of GAH is based on the reservoir controlled release multi particulate delivery system. Manufacturing process is reported to contain three stage coating process involves specialized equipment viz fluid bed processor. Various researchers attempted to prepare extended release formulations using Eudragit based matrix systems by preparing tablet as well as multi particulate dosage systems. Authors have studied matrix based mini tablets and reservoir based minitables approaches and found hydrophilic matrix based minitables approach as suitable.

Reference formulation of PHH is based on Geomatrix™ platform technology involving use bilayer compression machine. Other researcher's attempted preparing single layer delayed and controlled release systems based on hydrophilic polymers. Enhanced bioavailability was also reported for one such attempt as against the reference formulation. Authors have worked on different matrix and reservoir based systems, and HPMC-based matrix tablets which were further coated using methacrylic acid copolymer were found to be a suitable approach.

CONFLICT OF INTERESTS

The authors report no conflicts of interest. The authors are alone responsible for the content and writing of the paper.

REFERENCES

- Conte U, Maggi L, Colombo P, La MA. Multi-layered hydrophilic matrices as constant release devices (Geomatrix® systems). J Controlled Release 1993;26:39-47.
- Conte U, Maggi L. Modulation of the dissolution profiles from Geomatrix multi-layer matrix tablets containing drugs of different solubility. Biomaterials 1996;17:889-6.
- Rodriguez M, Vila-Jato JL, Torres D. Design of a new multi particulate system for potential site-specific and controlled drug delivery to the colonic region. J Controlled Release 1998;55:67-77.

4. Khan GM, Jiabi Z. Formulation and *in vitro* evaluation of Ibuprofen-Carbopol 974P-NF controlled release matrix tablets III: influence of co excipients on the release rate of the drug. J Controlled Release 1998;54:185-90.
5. Efentakis M, Koutlis A, Vlachou M. Development and evaluation of multiple oral unit and single unit hydrophilic controlled release systems. AAPS PharmSciTech 2000;4:1-9.
6. Verma RK, Garg S. Current status of drug delivery technologies and future directions. Pharm Technol 2011;25:1-4.
7. Krishnaiah YSR, Karthikeyan RS, Gourisankar V, Satyanarayana V. Three-layer guar gum matrix tablet formulations for oral controlled delivery of highly soluble Trimetazidine dihydrochloride. J Controlled Release 2002;81:45-56.
8. Tiwari SB, Murthy KT, Pai MR, Mehta PR, Chowdary PB. Controlled release formulation of Tramadol hydrochloride using hydrophilic and hydrophobic matrix system. AAPS PharmSciTech 2003;4:1-6.
9. Rao VM, Engh K, Qiu Y. The design of pH-independent controlled release matrix tablets for acidic drugs. Int J Pharm 2003;252:81-6.
10. Royce A, Li S, Weaver M, Shah U. *In vivo* and *in vitro* evaluation of three controlled release principles of 6-N-cyclohexyl-2-O-methyladenosine. J Controlled Release 2004;97:79-90.
11. Korhonen O, Kanerva H, Vidgren M, Urtti A, Ketolainen J. Evaluation of novel starch acetate diltiazem controlled release tablets in healthy human volunteers. J Controlled Release 2004;95:515-20.
12. Kim C. Controlled release from triple layer donut-shaped tablets with enteric polymers. AAPS PharmSciTech 2005;6:1-8.
13. Jamzad S, Fassih R. Development of a controlled release low dose class II drug Glipizide. Int J Pharm 2005;312:24-32.
14. Lopes CM, Lobo JSM, Costa P. Directly compressed mini matrix tablets containing Ibuprofen: preparation and evaluation of sustained release. Drug Dev Ind Pharm 2006;32:95-106.
15. Siepmann F, Siepmann J, Walther M, MacRae RJ, Bodmeier R. Polymer blends for controlled release coatings. J Controlled Release 2007;125:1-15.
16. Ishida M, Abe K, Hashizume M, Kawamura M. A novel approach to sustained pseudoephedrine release: Differentially coated mini-tablets in HPMC capsules. Int J Pharm 2008;359:46-52.
17. Dey NS, Majumdar S, Rao MEB. Multiparticulate drug delivery systems for controlled release. Trop J Pharm Res 2008;7:1067-75.
18. Baloglu E, Senyigit T. A design and evaluation of layered matrix tablet formulations of metoprolol tartrate. AAPS PharmSciTech 2010;11:563-73.
19. Aboelwafa AA, Basalious EB. Optimization and *in vivo* pharmacokinetic study of a novel controlled release Venlafaxine hydrochloride three-layered tablet. AAPS PharmSciTech 2010;11:1026-37.
20. Rao NGR, Hadi MA, Panchal HA. A novel approach to sustained Montelukast sodium release: Differentially coated mini-tablets in HPMC capsules. Int J Pharm Biomed Res 2011;2:90-7.
21. Mamani PL, Caro RR, Veiga MD. Matrix tablets: the effect of hydroxypropyl methylcellulose/anhydrous dibasic calcium phosphate ratio on the release rate of a water soluble drug through the gastrointestinal tract *in vitro* tests. AAPS PharmSciTech 2012;13:1073-83.
22. Mashkovsky MD, Kruglikova-Lvova RP. On the pharmacology of the new alkaloid galantamine. Farmakologia Toxicologia 1951;14:27-30.
23. Proskurnina NF, Yakovleva AP. Alkaloids of *Galanthus woronowi*. II Isolation of a new alkaloid. Zhurnal Obshchei Khimii 1952;22:1899-902.
24. Inventor Davis. The method of treating Alzheimer's disease. US Patent No. 4663318; 1987.
25. Cronin JR. The plant alkaloid Galantamine: approved as a drug; sold as a supplement. Alternative Complementary Ther 2004;7:380-3.
26. Sean C Sweetman. Martindale: The complete drug reference. Chicago, IL: Pharmaceutical Press; 2005.
27. US Patent 7160559. Inventor Mc Gee, Controlled release Galantamine composition; 2007.
28. Kays LA, Sileno AP, Brandt GC, Foerder CA, Quay SC. *In vitro* formulation optimization of intranasal galantamine leading to enhanced bioavailability and reduced emetic response *in vivo*. Int J Pharm 2007;335:138-46.
29. Caro VD, Giandalia G, Siragusa MG, Campisi G, Giannola LI. Galantamine delivery on buccal mucosa: permeation enhancement and design of matrix tablets. J Bioequivalence Bioavailability 2009;1:127-34.
30. Jagadeesh TR, Chary RBR. Development of a novel oral multi-particulate drug delivery system of galantamine hydrobromide. J Pharm Res 2011;4:77-9.
31. Biswal B, Sharma D. Design, Development and evaluation of galantamine hydrobromide bilayer sustained release tablet. Pharm Lett 2013;5:12-9.
32. Woo FY, Basri M, Masoumi HRF, Ahmad MB, Ismail M. Formulation optimization of galantamine hydrobromide loaded gel drug reservoirs in a transdermal patch for Alzheimer's disease. Int J Nanomed 2015;10:3879-86.
33. Ravi V, Kumar TMP, Rao NR. Investigation of kondagogu gum as a carrier to develop polymeric blend beads of galantamine hydrobromide. World J Pharm Pharm Sci 2015;4:1802-6.
34. Khatavkar UN, Shimpi SL, Kumar KJ, Deo KD. Controlled release reservoir mini tablets approach for controlling the drug release of galantamine hydrobromide. Pharm Dev Technol 2012;17:437-42.
35. Khatavkar UN, Shimpi SL, Kumar KJ, Deo KD. Development and *in vivo* evaluation of novel monolithic controlled release compositions of galantamine hydrobromide as against reservoir technology. Pharm Dev Technol 2013;18:1148-58.
36. Sweetman SC. Martindale-the complete drug reference. 34th ed. London: Pharmaceutical Press; 2005.
37. WO 2007/035816. Assignee: Dr Reddy's Laboratories) Paroxetine compositions; 2007.
38. Jin SJ, Yoo YH, Kim MS, Kim JS, Park JS, Hwang SJ. Paroxetine hydrochloride controlled release POLYOX matrix tablets: screening of formulation variables using Plackett-Burman screening design. Arch Pharm Res 2008;31:399-405.
39. Khatavkar UN, Shimpi SL, Kumar KJ, Deo KD. Development and comparative evaluation of *in vitro*, *in vivo* properties of novel controlled release compositions of paroxetine hydrochloride hemihydrate as against Geomatrix™ platform technology. Drug Dev Ind Pharm 2013;39:1175-86.
40. Vaidya N, N yak R, Benhar R, Narayanswamy VB. Design and evaluation of controlled release tablets of paroxetine hydrochloride. Int Res J Pharm 2013;4:84-62.

How to site this article

- Umesh Nandkumar Khatavkar, K Jayaram Kumar, Shamkant Laxman Shimpi. Novel approaches for the development of oral controlled release compositions of galantamine hydrobromide and paroxetine hydrochloride hemihydrate: a review. Int J Appl Pharm 2016;8(3):1-6