

IN VITRO EVALUATION OF ANTIOXIDANT CAPACITY OF ALGERIAN ORIGANUM PLANT BY SPECTROPHOTOMETRICAL AND ELECTROCHEMICAL ASSAYS

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ABSTRACT

Objective: The aim of the present study was to evaluate the antioxidant capacity and total phenolic of ethanolic extracts of two plants; *Origanum majorana* and *Origanum vulgare*.

Methods: Total phenolic was estimated by the Folin-Ciocalteu method. The antioxidant capacity was measured using the 2,2-diphenyl-1-picrylhydrazyl (DPPH) method for spectrophotometrical technique and cyclic voltammetry for electrochemical assay.

Results: According to the results the leaves extracts have very important values for polyphenols (266.86 mg GAE/g and 194.78 mg GAE/g) and high antioxidant activity; DPPH ($IC_{50} = 1.37$ mg/L and $IC_{50} = 1.53$ mg/L) for species *majorana*, and *vulgare* respectively. The antioxidant capacity of two species of *Origanum* was measured using ascorbic acid equivalent antioxidant capacity assays. The *O. majorana* extract showed the highest antioxidant capacity (292.97 mg/g) followed by *O. vulgare* extract (163.64 mg/g).

Conclusion: The results show that the antioxidant capacity expressed in terms of ascorbic acid equivalent antioxidant capacity obtained from electrochemical experiments (cyclic voltammetry) is higher than that obtained from spectrophotometrical experiments using DPPH. This outcome can be attributed to the overestimation of the total polyphenolic contents due to the interferences of other non-phenolic species.

Keywords: *Origanum majorana*, *Origanum vulgare*, DPPH, Voltammetry cyclic.

INTRODUCTION

Marjoram was formerly classified as coming from a sister genus of *Oregano*, but is now officially a species of *Oregano* itself [1]. In New Zealand the names are often used interchangeably, though *marjoram* (also known as sweet marjoram) differs from *oregano* in having a milder flavour.

Oregano is one of the most studied herbs, as it has shown consistently high levels of phenolics, antioxidant activity both had extremely high levels of phenolics as well as antioxidant activity. *Oregano* similarly ranked very highly in a number of studies over a range of different antioxidant assays, demonstrating its various modes of antioxidant activity [2-4]. A range of phenolic compounds has been identified in *Oregano* including *rosmarinic*, *caffeic*, *p-coumaric* acids and *caffeoyl* derivatives, the phenolic monoterpenes, *carvacrol*, *thymol*, flavonoids, luteolin, apigeninmyricetin and quercetin [5-10]. The aim of this work is to measure the *IN VITRO* antioxidant capacity of the ethanolic extracts of south Algerian *Origanum*. We used the following two assays systems (i) DPPH radical scavenging activity assay and (ii) cyclic voltammetry assay [11-14]. Total phenolic contents of ethanolic extract of *Origanum majorana* and *Origanum vulgare* were determined by standard colorimetric method.

MATERIALS AND METHODS

Vegetable matter

The two species of *Origanum* family *Lamiatae* were cultivated in the area of El-Oued, south of Algeria.

Instrument: UV-Visible spectrophotometer

(PRIM Advanced SCHOTT Instruments GmbH), centrifuge machine (SLW centryge Ultra-8TL), PGP301 potentiostat with voltmaster 4 version 7.08 software (radiometer analytical SAS), rotary evaporator (IKA evaporator RV 06-ML).

Ethanolic extracts

The powders of each plant material (10 g) was extract with 135 mL of ethanol absolute into the Soxhlet apparatus, and was extracted for

3 hours. The liquids extracts were filtered by Whatman. The filtrate was concentrated under reduced pressure at 40 °C by rotary evaporator (BUCHI R-210, Switzerland) to eliminate the ethanol, and stored in -4°C to give a crude extract yielding 0.8165 g for fresh leaves of *Origanum majorana* and 0.711 g for *Origanum vulgare*, diluted in ethanol and distilled water for next concentrations needed in this work.

Chemicals and reagents

Gallic acid, DPPH, anhydrous sodium sulfate sodium carbonate 20% (Na_2CO_3), Folin-Ciocalteu (F-C) reagent, ascorbic acid, ethanol (95%), were procured from Sigma-Aldrich Inc (Paris, France), all other reagents used were of analytical grade.

Determination of total phenolic

The total phenolic contents were determined by Folin-Ciocalteu colorimetric method [15]. Briefly, 100 μ L of sample (diluted solution) and 500 μ L of Folin-Ciocalteu reagent were pipetted into an Eppendorf tube. The contents were vortexed for 10 seconds. 2 mL of 20% (w/v) sodium carbonate solution was added to stop the reaction. The reaction mixture was incubated for 30 minutes at room temperature; the absorbance was measured at 760 nm. Gallic acid concentrations ranging from 0 to 30 mg/mL were prepared, and the calibration curve was obtained using a linear fit ($Y = 3.435X$, $R^2 = 0.992$). The samples were analyzed in duplicate. All results presented are means (\pm SD) and were analyzed in three replications.

Evaluation of antioxidant capacity by spectrophotometrical techniques

Using the free radical scavenging determination

The DPPH free radical scavenging activity of all extracts was measured according to the well-known DPPH test. The radical scavenging activity using free-radical DPPH assay determinate with the method described by Hatano et al [16] and Falleh et al [17]. Briefly, 100 μ L sample of various concentration of ethanolic extract of *Origanum majorana* (0.312, 0.104, 0.078 and 0.062 mg/mL, $R^2 = 0.938$, $Y = 5.028x + 32.69$) and 100 μ L sample of various

concentration of ethanolic extract of *Origanum vulgare* (1.1, 0.55, 0.36, 0.275 and 0.22 mg/mL, $R^2=0.999$, $Y=8.718x+26.91$) was added to 1 mL of a DPPH methanolic solution (4.9 mg DPPH in 50 mL methanol 100%). The mixture was vigorously shaken and left to stand in the dark for 30 minutes at room temperature. The antioxidant activity was then measured by the decrease in absorption at 517 nm using UV-Visible spectrophotometer and corresponds to the extract ability to reduce the radical DPPH* to the yellow-coloured diphenilpicryldrazine. The antiradical activity was expressed as IC_{50} ($\mu\text{g/mL}$), the antiradical dose required to cause 50% and calculated by the following equation:

$$\text{DPPH scavenging activity (\%)} = \frac{(A_0 - A_1)}{A_0} \times 100(1)$$

Where A_0 is the absorbance of control at 30 minutes, A_1 is the absorbance of the sample extract at 30 minutes. All results presented are means (\pm SD) and were analyzed in three replications.

Evaluation of antioxidant capacity by electrochemical techniques

The measurement of the antioxidant capacity of the studied samples of *Origanum* was performed using an electrochemical method based on cyclic voltammetry techniques. Cyclic voltammetry measurements were performed in an electrochemical cell with a volumetric capacity of 50 mL containing a glassy carbon electrode (GCE) working electrode (radio meter analytical SAS), a Pt wire counter electrode and an Hg/Hg₂Cl₂ reference electrode (saturated with KCl). The potential was swept in direct scanning mode starting from -200 to +1000 mV with a scanning rate of 100 mV/s. To avoid reducing the sensitivity of the working electrode, the latter was polished after each cycle by rubbing its surface using alumina oxide (particle size 0.3 μm) before every electrochemical assay. After polishing it was rinsed thoroughly with bi distilled water for 30 seconds. The antioxidant capacity of the studied samples of *Origanum* was obtained using the current density of the anodic curve of the voltammogram. The calibration graph is obtained by plotting the current density of the anodic curve of the voltammogram of each sample of the standard versus its concentration. Ascorbic acid was used as a standard in the calculation of antioxidant capacity of the studied sample of *Origanum* because of its wide spreading in nature and also because its anodic current density displays excellent linearity toward ascorbic acid concentrations [18-19].

RESULTS AND DISCUSSION

Total phenolic contents in the selected plant

The total phenolic contents of *Origanum* species were measured using F-Creagent method.

These results obtained by Soxhlet extraction using ethanol absolute solvent are presented in table 1.

Table 1: Total polyphenol of ethanolic leaves extracts of genus *Origanum*.

Plant species	Polyphenols (mg GAE/g)
<i>Origanum majorana</i>	266.86 \pm 1.37
<i>Origanum vulgare</i>	194.78 \pm 1.49

Data are expressed as means \pm standard deviation of triplicate samples, Values with different row are significantly ($P < 0.05$).

As can be seen from the table 1, significant phenolic contents were observed for different ethanolic extracts of *Origanum majorana* (266.86 mg GAE/g) and *Origanum vulgare* (194.78 mg GAE/g). These concentrations significantly higher if are compared to other medicinal plants like *G. multifloral* 12.36 mg GAE/g and *G. villosa* 20.81 mg GAE/g [20], 70.07 mg GAE/g DW for *M. edule* [17]. According to Zheng & Wang [21], two *Oregano* species tested (*Origanum vulgare* and *Origanum majorana*) both had extremely high levels of phenolic as well as antioxidant activity.

Free radical DPPH scavenging assay

The DPPH radical scavenging activity of ethanolic extract leaves of the two species of *Origanum* presented in table 2. For ethanolic extract of *O. majorana* obtained the higher value ($IC_{50}=1.37 \pm 0.08$ mg/L), the value found in *Origanum vulgare* ($IC_{50}=1.53 \pm 0.07$ mg/L). The antioxidant capacity of the two species of *Origanum* is higher than the positive control BHA ($IC_{50}=28.27 \pm 3.85$ mg/L), this antioxidant capacity free radical scavenger DPPH related with the quantity of total polyphenol composition [22]. The relationship is related to their ability to antioxidant activity, free radical scavenger [23]. Similar results were observed in relation to lard [24-25]. The IC_{50} values are inversely proportional to the anti-radical activity. The values of all ARP (Anti radical activity, $ARP=1/IC_{50}$) [26] extracts are significant, moreover, these values do not tend and away from zero. The more ARP increases; we can say that our extracts have antioxidant activity. All IC_{50} are very low ranging between 1.37 $\mu\text{g/mL}$ and 28.7 $\mu\text{g/mL}$, under this setting sequestration, capacity radical are listed in order:

Origanum majorana > *Origanum vulgare* > atocopherol > BHA

Table 2: DPPH radical scavenging activity (IC_{50} in $\mu\text{g/mL}$) of the two extracts and ARP authentic standards

Extracts and standards	DPPH test (IC_{50} in $\mu\text{g/mL}$)	ARP*
<i>Origanum majorana</i>	1.37 \pm 0.08	0.072
<i>Origanum vulgare</i>	1.53 \pm 0.07	0.065
BHA[27]	28.27 \pm 3.85	0.035
Atocopherol[27]	15.99 \pm 0.25	0.062

Data are expressed as means \pm standard deviation of triplicate samples, Values with different row are significantly ($P < 0.05$). * anti-radical activity

Cyclic voltammetry assay

In order to express the antioxidant capacity of different species of the *Origanum* extracts in equivalent terms of ascorbic acid equivalent antioxidant capacity, different concentrations of the standards ascorbic acid (0.049 to 0.868 g/L) were plotted versus the anodic current density obtained from different cyclic voltammograms at pH 7 in 0.2 M phosphate buffer solution as a supporting electrolyte using a 3 mm-diameter glassy carbon electrode present typical irreversible oxidation processes with the existence of an irreversible one oxidation peak at 260mV (figure 1). As can be seen there is an increase in peak current with the increase in ascorbic acid concentrations which leads to a linear relation between these two parameters [28].

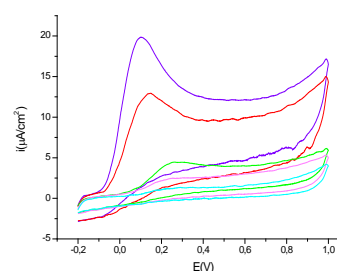


Fig. 1: Cyclic voltammogram referring to different ascorbic acid concentrations

The equation obtained from the linear calibration graph in the studied concentration range for ascorbic acid is, $y = 127.39x + 0.12$ (where y represents the value of the anodic current density and x , the value of standards concentration, expressed as g/L), with a correlation coefficient of $r^2 = 0.998$. The total antioxidant capacity was calculated based on the following equation:

$$TAC = \frac{C_1}{C_2} (2)$$

Where TAC is total antioxidant capacity, C_1 is the *Origanum* sample extract concentration (g/mL), C_2 is the sample concentration in the electrochemical cell (g/mL) calculated by replacing the current density obtained from different voltammograms of sample extracts in the equation $y = 127.39x + 0.12$. Antioxidant capacity of two *Origanum* varieties from [5, 7] showed higher antioxidant capacity.

Evaluation of antioxidant capacity of *Origanummajorana*

As it can be seen from voltammogram of figure 2, the ethanolic extract of *Origanummajorana* do not response in the same manner as the standard ascorbic acid. The voltammogram of extract represent two peaks for oxidation at 239.9 mV for first oxidation and another for second oxidation at 674.01 mV. This irreversible electrochemical behavior of ethanolic extract may indicate that the *O. majorana* extract contain a different polyphenolic contents of that of the standard ascorbic acid. Electrochemical data calculated from

voltammetric measurements of voltammogram of figure 2 are presented in table 3.

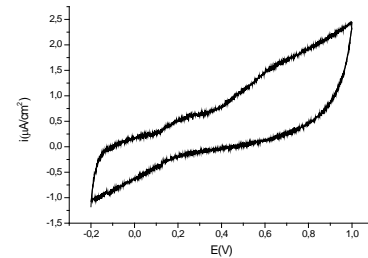


Fig. 2: Cyclic voltammogram of *Origanummajorana* ethanolic extract in pH 7, 0.2M phosphate buffer solution at scan rate 100 mV/s.

Table 3: Electrochemical data of ethanolic extract extracts of *O. majorana*

Ethanolic extract	Epa (mV)*	ipa* ($\mu\text{A}/\text{cm}^2$)	Epa** (mV)	ipa** ($\mu\text{A}/\text{cm}^2$)
<i>O. majorana</i>	239.9	0.5504	674.01	1.4406

*First oxidation **second oxidation

Evaluation of antioxidant capacity of *Origanumvulgare*

Figure 3 indicate that ethanolic extract of *Origanumvulgare* present the same irreversible electrochemical behavior of that of the standard ascorbic acid, although with oxidation potential value is less positive than ascorbic acid, around 13,4 to 212 mV. This may indicate that, under this electrochemical conditions, the extract of *O. vulgre* contain the same polyphenolic contents of the standards ascorbic acid. Electrochemical data obtained from voltammogram of ethanolic extract of *O. vulgre* sample is summarized in table 4.

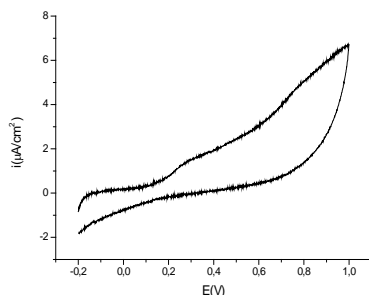


Fig. 3: Cyclic voltammogram of *Origanumvulgare* ethanolic extract in pH 7, 0.2M phosphate buffer solution at scan rate 100 mV/s.

Table 4: Electrochemical data of ethanolic extract of *O. vulgare*

Ethanolic extract	Epa (mV)	ipa* ($\mu\text{A}/\text{cm}^2$)
<i>O. vulgare</i>	212	0.8613

The ascorbic acid equivalent antioxidant capacity of different species of the two studied samples extracts calculated from the calibration graphs is summarized in table 5.

Table 5: Antioxidant capacities of two different varieties of *Origanum*

variety	Ascorbic acid antioxidant capacities	
	Ethanolic extracts (mg/g)	
<i>O. majorana</i>	262.97	
<i>O. vulgare</i>	163.64	

The ascorbic acid equivalent antioxidant capacity values for different variety of ethanolic extract of studied *Origanum* indicate that *Origanummajorana* variety was the most effective with the highest ascorbic acid equivalent antioxidant capacity value (262.97 mg/g), followed by *Origanumvulgare* variety, ascorbic acid equivalent antioxidant capacity value (163.64mg/g).

CONCLUSION

Both the spectrophotometrical (DPPH) and electrochemical (ascorbic acid equivalent antioxidant capacity) assays suggest that ethanolic extract of two species of *Origanum* shows *IN VITRO* antioxidant activities by inhibiting DPPH which may be due to presence phenolic compounds found in preliminary phytochemical screening. The results also show that the antioxidant capacity of *Origanummajorana* expressed in terms of ascorbic acid equivalent antioxidant capacity (AEAC) is higher than that obtained for *Origanumvulgare*. Also the results show that the antioxidant capacity expressed in terms of ascorbic acid equivalent antioxidant capacity obtained from electrochemical experiments (cyclic voltammetry) is higher than that obtained from spectrophotometrical experiments using DPPH. This outcome can be attributed to the overestimation of the total polyphenolic contents due to the interferences of other non-phenolic species.

CONFLICT OF INTERESTS

Declared None

REFERENCES

- McGee H. On food and cooking. Revised ed. New York: Scribner; 2004. p. 884.
- Capecka E, Mareczek A, Leja M. Antioxidant activity of fresh and dry herbs of some *Lamiaceae* species. Food Chem 2005;93(2):223-6.
- Dragland S, Senoo H, Wake K, Holte K, Blomhoff R. Several culinary and medicinal herbs are important sources of dietary antioxidants. J Nut 2003;133(5):1286-90.
- Shan B, Cai YZ, Sun M, Corke H. Antioxidant capacity of 26 spices extracts and characterization of their phenolic constituents. J Agricultural Food Chem 2005;53(20):7749-59.
- Yanishlieva NV, Marinova E, Pokorný J. Natural antioxidants from herbs and spices. Eur J Lipid Sci Technol 2006;108(9):776-93.
- Vichi S, Zitterl-Eglseer KMJ, Franz C. Determination of the presence of antioxidants deriving from *sage* and *oregano* extracts added to animal fat by means of assessment of theradical scavenging capacity by photochemiluminescence analysis. Nahrung/Food 2001;45(2):101-4.

7. Pellegrini N, Serafini M, Salvatore S, Del Rio D, Bianchi M, Brighenti F. Totalantioxidant capacity of spices, dried fruits, nuts, pulses, cereals and sweets consumed in Italy assessed by three different *IN VITRO* assays. *Mol Nutr Food Res* 2006;50(11):1030-8.
8. Beddows CG, Jagait C, Kelly MJ. Preservation of alpha-tocopherol in sunflower oil by herbs and spices. *Int J Food Sci Nutr* 2000;51(5):327-39.
9. Benchikha N. Etude approfondie de quelques huiles essentielles en vue de leur utilisation dans l'industrie pharmaceutique, PHD thesis es-sciences, Univ. Biskra. Algeria; 2010.
10. Benchikha N. Extraction and antioxidant activities of two species *origanum* plant containing phenolic and flavonoid compounds. *Fund App Sci* 2013;5(1):20-128.
11. Rebiai A, Lanez T, Belfar ML. *Int J Pharmacol* 2011;7(1):113-8.
12. Campanella L, Favero G, Persi L, Tomassetti M. *J Pharm Biomed Anal* 2001;24:1055-64.
13. Cosio MS, Buratti S, Mannino S, Denedetti S. *Food Chem* 2006;97:725-31.
14. Saha MA, Alia M, Rebiai A, Lanez T. *J Fun App Sci* 2011;3(2):35-43.
15. Moreira L, Dias LG, Pereira JA. Antioxidant properties, total phenols and pollen analysis of propolis samples from Portugal. *Food Chem Toxicol* 2008;46:3482-5.
16. Hatano T, Kagaw H, Yasuhara T. Two new flavonoid and other constituents in *licorice* root: their relative astringency and radical scavenging effects. *Chem Pharm Bull* 1989;36:2090-7.
17. Falleh F, Ksouri K, Oueslati S. Interspecific variability of antioxidant activities and phenolic composition in *Mesembryanthemum* genus. *Food Chem Toxicol* 2009;47:2308-13.
18. Laskar RA, Roy IN, Begum NA. *Food Chem* 2010;122:233-7.
19. Oyaizu M. Studies on product of browning reaction prepared from glucose amine. *Jpn J Nutr* 1986;44:307-15.
20. Liu HY, Qiu NX, Ding H. Polyphenols contents and antioxidant capacity of 68 Chinese herbals suitable for medical or food uses. *Food Res Int* 2008;41:363-70.
21. Zheng W, Wang SY. Antioxidant activity and phenolic compounds in selected herbs. *J Agricultural Food Chem* 2001;49(11):5165-70.
22. Julia V, Mario R, Maria Cecili L. Polyphenol input to the antioxidant activity of yerbamate (*Ilex paraguariensis*) extracts. *LWT-Food Sci Technol* 2012;45:28-35.
23. Neha B, Harinder SO, Dewinder SU. Total phenolic content and antioxidant capacity of extracts obtained from six important fruit residues. *Food Res Int* 2011;44:391-6.
24. Vichi S, Zitterl-Eglseer KM J, Franz C. Determination of the presence of antioxidants deriving from sage and oregano extracts added to animal fat by means of assessment of the radical scavenging capacity by photochemiluminescence analysis. *Nahrung/Food* 2001;45(2):101-4.
25. Castilho PC, Savluchinske-Feio S, Weinhold TS, Gouveia SC. Evaluation of the antimicrobial and antioxidant activities of essential oils, extracts and their main components from *oregano* from Madeira Island, Portugal. *Food Control* 2012;23:552-8.
26. Markowicz Bastos DH, Saldanha LA, Catharino RR, Sawaya ACHF, Cunhal BS, Carvalho PO, *et al.* Phenolic Antioxidants Identified by ESI-MS from Yerba Maté (*Ilex paraguariensis*) and Green Tea (*Cameliasinensis*) Extracts. *Mol* 2007;12:423-32.
27. Lanez T, Rebiai A. Development of an electrochemical method for the measurement of antioxidant capacity of pure compounds and natural substances extracts, ILCPA; 2013. p. 51.
28. Djouadi A. Evaluation de l'activité antioxydante des polyphénols extraits de deux variétés de *Solanum melongena L.* de la région d'El-Oued par voltampérométrie cyclique et ondes carrées, academic master, Univ. of El-oued, Algeria; 2012.