

Original Article

INHIBITION OF UREASE ENZYME ACTIVITY BY UREA AND THIOUREA DERIVATIVES OF DIPEPTIDES CONJUGATED 2, 3-DICHLOROPHENYL PIPERAZINE

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ABSTRACT

Objective: Studies on enzyme inhibition remain an important area of pharmaceutical research since these studies have led to the discoveries of drugs used in a variety of physiological conditions. In search of novel urease enzyme inhibitors, four dipeptides were synthesized, conjugated to 2,3-dichlorophenyl piperazine analogue and converted into urea/thiourea derivatives.

Methods: The peptides were synthesized by solution phase method and conjugated to 2,3-dichlorophenyl piperazine (PZN) using 1-ethyl-3-(3-dimethyl aminopropyl) carbodiimide (EDCI)/hydroxybenzotriazole (HOBt) as a coupling agent and *N*-methylmorpholine (NMM) as a base. The tert-butyloxycarbonyl (Boc) group was removed using trifluoroacetic acid (TFA), neutralized with NMM and converted to urea and thiourea derivatives using substituted phenyl isocyanates and isothiocyanates respectively.

Results: Most of the synthesized analogues were found to be good inhibitors of urease enzyme activity. The conjugates of thiourea with F and Cl substituents at *meta* or *para* positions showed predominant urease inhibitory activity. The analogue 23 was nearly 10 fold (2 μ M) more potent than the reference standard, 21.0 \pm 0.11 μ M.

Conclusion: The reported activity was correlated with some of the literature reported urease inhibitors and found that our compound 23 was more potent than the existing ones.

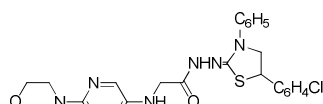
Keywords: Amino acids, Piperazine, Conjugation, Urea/thiourea, Urease inhibitors

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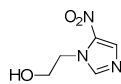
INTRODUCTION

Activity of urease (E. C 3.5.1.5) has been shown to be an important virulence determinant in the pathogenesis of many clinical conditions, which are detrimental for human and animal health as well as for agriculture. Urease is directly involved in the formation of infectious stones and contributes to the pathogenesis of urolithiasis, pyelonephritis, ammonia and hepatic encephalopathy, hepatic coma, urinary catheter encrustation [1-3]. It is also known to be a major cause of pathologies induced by *Helicobacter pylori* (HP), which allows bacteria to survive at low pH of the stomach during colonization and therefore plays an important role in the pathogenesis of gastric and peptic ulcer [2]. In agriculture, high urease activity causes significant environmental and economic problems by releasing abnormally large amounts of ammonia into

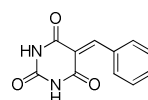
the atmosphere during urea fertilization. This further induces plant damage primarily by depriving them from their essential nutrient, and secondly ammonia toxicity and increase the pH of the soil [1, 2]. Therefore, strategies based on urease inhibition were considered as the first line of treatment for infections caused by urease producing bacteria. Several classes of molecules have been tested as urease inhibitors (fig. 1) [4-9], but part of them was prevented from using *in vivo* because of their toxicity or instability. As a consequence agents currently used to inhibit urease have some deleterious side effects. Hence, the present work was intended to search for better compounds which would enable to replace the existing ones suffering from serious limitations. Meanwhile, the studies on novel urease inhibitors are essential not only for the basic research on urease biochemistry but also for the possible development of a highly needed therapy for urease mediated bacterial infections.



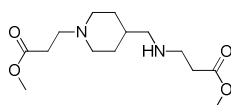
Morpholine = 2.37 μ M



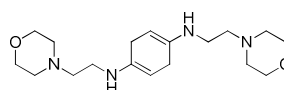
Metronidazole = 12 μ M



Barbiturates = 13 μ M



Substituted piperidine = 19.38 μ M



Benzoquinone derivative = 27.30 μ M

Urea is a common structural feature of many biologically active compounds such as enzyme inhibitors and pseudo peptides. The

literature on urease inhibitors with possible applications in medicine showed urea derivatives and heterocyclic compounds

constitute the major classes of structures with such activity. These ureas/thioureas display a wide range of biological activities including antibacterial, antifungal, antitubercular, antithyroid, anthelmintic, antidiabetic, anticancer, insecticidal, herbicidal and antimalarial properties [10-14].

Piperazine scaffold has been considered as a moiety possessing a myriad spectrum of medicinal properties. Diversity of biological responses profile has attracted the considerable interest of several researchers across the globe to explore this skeleton for its assorted therapeutic significance. Various novel classes of structurally different piperazine motifs have been designed and synthesized depicting potential interventions and versatile type of biological activities such as antiurease, antibacterial, antidiabetic, anti-inflammatory, anticoagulant and also found to have used in acute toxicity study [15-19]. There is a large amount of literature dealing with the physiological significance of amino acids. Their effects on a variety of metabolic and physiological ramifications are among the most investigated topics in biomedical sciences. In addition, taking into consideration the need of more urease inhibitors and afore mentioned biological significance of piperazine, ureas/thioureas, amino acids functions and coupled with our research focused on the development of novel potential bioactive agents [20-23]. The aim of this report was to develop some new antiurease agents with better therapeutic potential.

MATERIALS AND METHODS

General

All amino acids used except glycine were of *L*-configuration unless otherwise mentioned. All Boc-amino acids, EDCI, HOBt and TFA were purchased from Advanced Chem. Tech. (Louisville, Kentucky, USA). Isobutyl chloroformate (IBCF) and NMM were purchased from Sigma Chemical Co. (St. Louis, MO). All solvents and reagents used for the synthesis were of analytical grade. The progress of the reaction was monitored by thin layer chromatography (TLC) using silica gel coated on glass plates with the solvent system comprising chloroform/methanol/acetic acid in the ratio 98:2:3 (R_f^a) and 95:5:3 (R_f^b) throughout the study and the compounds on TLC plates were detected by iodine vapors. Melting points (MP) were determined on a Superfit melting point apparatus (India) and are uncorrected. Infrared (IR) spectra of the compounds were recorded on Jasco Spectrometer (USA). ^1H nuclear magnetic resonance (NMR) spectra were obtained on VARIAN 400 MHz instrument (USA) using DMSO- d_6 and the chemical shifts are reported as parts per million (δ ppm) using TMS as an internal standard. Mass spectra were obtained on Bruker (model HP-1100) (USA) electrospray mass spectrometer. Elemental analysis was performed by using VARIO EL III Elementar (Germany). Jack bean urease enzyme for urease inhibition studies was purchased from Research Organics, Cleveland, while other chemicals were purchased from Sigma Aldrich.

Synthesis

Synthesis of Boc-Xaa-Pro-OBzl where Xaa = Phe (I), Val (II), Tyr(2,6-Cl₂Bzl) (III) and Lys(Z) (IV)

To a Boc-Xaa-OH (0.005 mol) dissolved in acetonitrile (10 ml/g) and cooled to 0 °C and then added NMM (1.10 ml, 0.005 mol). The solution was cooled to -15 °C \pm 1 °C and then IBCF (0.77 ml, 0.005 mol) was added under stirring maintaining the temperature at -15 °C. After stirring the reaction mixture for 10 min a precooled solution of HCl-Pro-OBzl (1.209 g) in DMF (12 ml) and NMM (1.09 ml, 0.005 mol) was poured slowly. After 20 min the pH of the solution was adjusted to 8 by adding NMM and the reaction mixture was stirred overnight at room temperature.

Acetonitrile was removed under reduced pressure and the residual DMF solution was poured into about 200 ml ice cold 90 % saturated KHCO₃ solution and stirred for 30 min. The product precipitated was extracted into CHCl₃. The organic layer was washed with 5 % NaHCO₃ solution (3 \times 20 ml), water (2 \times 20 ml) and 0.1 N HCl solution (3 \times 20 ml) followed by brine (2 \times 20 ml). The chloroform layer was dried over anhydrous Na₂SO₄. The solvent was removed under reduced pressure and triturated with dry ether or petroleum ether to obtain the products I-IV.

Synthesis of Boc-Xaa-Pro-OH

Each dipeptide (I-IV) (0.004 mol) was dissolved in methanol (10 ml/g) and 1N NaOH (2 eq) was added and stirred for 2 h at rt. The completion of the reaction was monitored by TLC and the solvent was evaporated, cooled, neutralized with cold 1 N HCl and extracted with CHCl₃. The organic layer was washed with 1 N HCl followed by H₂O and dried over anhydrous Na₂SO₄. The solvent was removed under pressure and triturated with ether, filtered, washed with ether and dried to obtain debenzylated peptides (V-VIII).

Boc-Phe-Pro-OH (V)

Yield = 92.0%; R_f^a = 0.61; R_f^b = 0.75; M. P. = 97-100 °C (Lit. 100 °C) [24]; ^1H NMR (DMSO- d_6 , δ ppm): Boc = 1.41 (9H, s); Phe = 3.42-3.53 (2H, d, $-\beta\text{CH}_2$); 4.86-4.91 (1H, m, $-\alpha\text{CH}$); 6.54-7.40 (5H, m, Ar-H); Pro = 3.92-3.97 (1H, t, $-\alpha\text{CH}$), 1.68-2.22 (4H, m, $-\beta\gamma\text{CH}_2$), 3.40-3.51 (2H, m, $-\delta\text{CH}_2$); Mass = 367.40 ($M^+ + 1$); Elem. Anal. = calcd. for C₁₉H₂₆N₂O₅: C: 62.97; H: 7.23; N: 7.73; Found: C: 62.95; H: 7.20; N: 7.73

Boc-Val-Pro-OH (VI)

Yield = 94.0%; R_f^a = 0.56; R_f^b = 0.68; M. P. = 80-82 °C (Lit. 83 °C) [25]; ^1H NMR (DMSO- d_6 , δ ppm): Boc = 1.42 (9H, s); Val = 1.03 (6H, d, (CH₃)₂), 1.88-1.90 (1H, m, $-\beta\text{CH}$), 3.53-3.64 (1H, m, $-\alpha\text{CH}$); Pro = 3.69-3.73 (1H, t, $-\alpha\text{CH}$), 1.90-2.35 (4H, m, $-\beta\gamma\text{CH}_2$), 2.97-3.52 (2H, m, $-\delta\text{CH}_2$); Mass = 315.30 ($M^+ + 1$); Elem. Anal. = calcd. for C₁₅H₂₂N₂O₅: C: 57.31; H: 8.34; N: 8.91; Found: C: 57.32; H: 8.32; N: 8.88

Boc-Tyr(2,6-Cl₂Bzl)-Pro-OH (VII)

Yield = 89.2%; R_f^a = 0.56; R_f^b = 0.69; M. P. = 95-98 °C; ^1H NMR (DMSO- d_6 , δ ppm): Boc = 1.43 (9H, s); Tyr = 3.66-3.76 (2H, d, $-\beta\text{CH}_2$); 4.81-4.87 (1H, t, $-\alpha\text{CH}$); 5.17 (2H, s, side chain-CH₂); 6.70-7.90 (7H, m, ArH); Pro = 3.35-3.48 (1H, t, $-\alpha\text{CH}$), 1.68-1.88 (4H, m, $-\beta\gamma\text{CH}_2$), 2.91-3.13 (2H, m, $-\delta\text{CH}_2$); Mass = 538.41 ($M^+ + 1$); Elem. Anal. = calcd. for C₂₆H₃₀Cl₂N₂O₆: C: 58.11; H: 5.63; N: 5.21; Found: C: 58.12; H: 5.62; N: 5.28

Boc-Lys(Z)-Pro-OH (VIII)

Yield = 89.2%; R_f^a = 0.56; R_f^b = 0.64; M. P. = Gum; ^1H NMR (DMSO- d_6 , δ ppm): Boc = 1.43 (9H, s); Lys = 1.70-1.72 (2H, m, $-\delta\text{CH}_2$); 1.85-1.91 (2H, m, $-\gamma\text{CH}_2$); 1.93-2.01 (2H, m, $-\beta\text{CH}_2$); 3.18-3.25 (2H, m, $-\alpha\text{CH}_2$); 4.06 (1H, m, $-\alpha\text{CH}$); 8.13 (1H, s, -NH); 5.15 (2H, s, benzyl-CH₂); 7.18-7.43 (5H, m, ArH); Pro = 4.05-4.10 (1H, t, $-\alpha\text{CH}$), 1.67-2.25 (4H, m, $-\beta\gamma\text{CH}_2$), 3.72-3.78 (2H, m, $-\delta\text{CH}_2$); Mass = 478.51 ($M^+ + 1$); Elem. Anal. = calcd. for C₂₄H₃₅N₃O₇: C: 60.36; H: 7.39; N: 8.80; Found: C: 60.32; H: 7.32; N: 8.88

General procedure for the coupling of piperazine derivative with peptides (1-4)

PZN was synthesized as previously reported method [26]. To Boc-Xaa-Pro-OH (0.01 mol) dissolved in acetonitrile (10 ml/g of compound) and cooled to 0 °C was added NMM (1.10 ml, 0.01 mol). To this EDCI (1.917 g, 0.01 mol) was added and stirred while maintaining the temperature at 0 °C. After stirring the reaction mixture for 10 min at this temperature, HOBt (1.531 g, 0.01 mol) in dimethyl formamide (DMF) (15 ml) was added slowly. The reaction mixture was stirred for an additional 10 min and a pre-cooled solution of PZN (2.68 g, 0.01 mol) and NMM (1.10 ml, 0.01 mol) in DMF (25 ml) was added slowly. After 20 min, pH of the solution was adjusted to 8 by the addition of NMM and the reaction mixture was stirred overnight at room temperature. Acetonitrile was removed under reduced pressure and the residual DMF was poured into about 500 ml ice-cold 90% saturated potassium bicarbonate (KHCO₃) solution and stirred for 30 min. The precipitated compound was extracted into chloroform and washed sequentially with 5% sodium bicarbonate (NaHCO₃) solution (3 \times 20 ml), water (3 \times 20 ml), 0.1N cold HCl (3 \times 20 ml) followed by brine. The organic layer was dried over anhydrous Na₂SO₄, the solvent was removed under reduced pressure, triturated with ether, filtered and dried to get the conjugates 1-4.

tert-butyl (1-(2-(4-(2,3-dichlorophenyl)piperazine-1-carbonyl)pyrrolidin-1-yl)-1-oxo-3-phenylpropan-2-yl)carbamate (1)

Yield = 90.0%; R_f^a = 0.58; R_f^b = 0.65; M. P. = Gum; ^1H NMR (DMSO- d_6 , δ ppm): Boc = 1.43 (9H, s); Phe = 3.41-3.51 (2H, d, $-\beta\text{CH}_2$), 4.59-4.91 (1H, m, $-\alpha\text{CH}$), 6.52-7.40 (5H, m, Ar-H); Pro = 3.79-3.94 (1H, t, $-\alpha\text{CH}$),

1.68-2.20 (4H, m, $-\beta\gamma\text{CH}_2$), 3.38-3.49 (2H, m, $-\delta\text{CH}_2$); PZN = 3.26-3.37 (4H, m, $-\text{CH}_2$); 3.42-3.46 (4H, m, $-\text{CH}_2$); 6.52-7.40 (3H, m, Ar-H). Mass = 576.48 ($M^+ + 1$); Elem. Anal. = calcd. for $\text{C}_{29}\text{H}_{36}\text{N}_4\text{O}_4$: C: 60.52; H: 6.30; N: 9.73; Found: C: 60.55; H: 6.31; N: 9.75

tert-butyl (1-(2-(4-(2,3-dichlorophenyl)piperazine-1-carbonyl)pyrrolidin-1-yl)-3-methyl-1-oxobutan-2-yl)carbamate (2)

Yield = 88.8%; $R^a = 0.48$; $R^b = 0.57$; M. P. = Gum; $^1\text{H NMR}$ (DMSO- d_6 , δ ppm): Boc = 1.43 (9H, s); Val = 1.06 (6H, d, $-(\text{CH}_3)_2$), 1.88-1.91 (1H, m, $-\beta\text{CH}$), 4.34 (1H, s, $-\alpha\text{CH}$); Pro = 4.02 (1H, t, $-\alpha\text{CH}$), 1.68-2.24 (4H, m, $-\beta\gamma\text{CH}_2$), 3.74-3.78 (2H, m, $-\delta\text{CH}_2$); PZN = 3.34-3.42 (4H, m, $-\text{CH}_2$); 3.47-3.50 (4H, m, $-\text{CH}_2$); 7.14-7.28 (3H, m, Ar-H). Mass = 528.45 ($M^+ + 1$); Elem. Anal. = calcd. for $\text{C}_{25}\text{H}_{36}\text{Cl}_2\text{N}_4\text{O}_4$: C: 56.92; H: 6.88; N: 10.62; Found: C: 56.90; H: 6.82; N: 10.68

tert-butyl (3-(4-((2,6-dichlorobenzoyloxy)phenyl)-1-(2-(4-(2,3-dichlorophenyl)piperazine-1-carbonyl)pyrrolidin-1-yl)-1-oxopropan-2-yl)carbamate (3)

Yield = 86.2%; $R^a = 0.56$; $R^b = 0.68$; M. P. = Gum; $^1\text{H NMR}$ (DMSO- d_6 , δ ppm): Boc = 1.43 (9H, s); Tyr = 3.72 (2H, d, $-\beta\text{CH}_2$); 4.81-4.89 (1H, m, $-\alpha\text{CH}$); 5.15 (2H, s, side chain- CH_2); 6.98-7.92 (7H, m, ArH); Pro = 3.48 (1H, t, $-\alpha\text{CH}$), 1.68-1.88 (4H, m, $-\beta\gamma\text{CH}_2$), 2.94-3.16 (2H, m, $-\delta\text{CH}_2$); PZN = 3.36-3.48 (4H, m, $-\text{CH}_2$); 3.62-3.70 (4H, m, $-\text{CH}_2$); 6.98-7.92 (3H, m, Ar-H). Mass = 751.52 ($M^+ + 1$); Elem. Anal. = calcd. for $\text{C}_{36}\text{H}_{40}\text{Cl}_4\text{N}_4\text{O}_5$: C: 57.61; H: 5.37; N: 7.46; Found: C: 57.62; H: 5.32; N: 7.48

benzyl tert-butyl (6-(2-(4-(2,3-dichlorophenyl)piperazine-1-carbonyl)pyrrolidin-1-yl)-6-oxohexane-1,5-diyl)dicarbamate (4)

Yield = 89.2%; $R^a = 0.56$; $R^b = 0.64$; M. P. = Gum; $^1\text{H NMR}$ (DMSO- d_6 , δ ppm): Boc = 1.43 (9H, s); Lys = 1.70-1.72 (2H, m, $-\delta\text{CH}_2$), 1.85-1.91 (2H, m, $-\gamma\text{CH}_2$), 1.93-2.01 (2H, m, $-\beta\text{CH}_2$), 3.18-3.25 (2H, m, $-\epsilon\text{CH}_2$); 4.06 (1H, m, $-\alpha\text{CH}$); 8.13 (1H, s, $-\text{NH}$); 5.15 (2H, s, benzyl- CH_2); 7.18-7.43 (5H, m, ArH); Pro = 4.05 (1H, t, $-\alpha\text{CH}$), 1.65-2.21 (4H, m, $-\beta\gamma\text{CH}_2$), 3.71-3.76 (2H, m, $-\delta\text{CH}_2$); PZN = 3.34-3.42 (4H, m, $-\text{CH}_2$); 3.47-3.50 (4H, m, $-\text{CH}_2$); 7.14-7.28 (3H, m, Ar-H). Mass = 691.65 ($M^+ + 1$); Elem. Anal. = calcd. for $\text{C}_{34}\text{H}_{45}\text{Cl}_2\text{N}_5\text{O}_6$: C: 59.13; H: 6.57; N: 10.14; Found: C: 59.12; H: 6.52; N: 10.18

General procedure for the synthesis of ureido and thioureido derivatives (5-36)

Each time, Boc-Xaa-Pro-PZN (0.15g) was stirred with 1.5 ml of TFA for 45 min at room temperature. After completion of the reaction (monitored by TLC), TFA was removed under vacuum, triturated with dry ether, filtered, washed with ether and dried to obtain TFA. H-Xaa-Pro-PZN.

Further, TFA. H-Xaa-Pro-PZN (0.001 mol) was dissolved in DMF (10 ml/g of the compound), cooled to 0 °C and NMM (0.10 ml, 0.001 mol) was added. To this solution respective substituted phenyl isocyanates and isothiocyanates (0.0012 mol) was added drop-wise

while maintaining the temperature at 0 °C. The reaction mixture was stirred for 8h slowly warming to room temperature. DMF was evaporated under high vacuum and the residue was poured into about 50 ml ice-cold 90% saturated KHCO_3 solution and stirred for 30 min. The precipitate was extracted into chloroform and washed sequentially with 5% NaHCO_3 solution (2×10 ml), water (2×10 ml), 0.1N citric acid (2×10 ml) followed by brine. The organic layer was dried over anhydrous Na_2SO_4 . The solvent was removed under reduced pressure, triturated with hexane filtered and dried under vacuum to obtain urea/thiourea derivatives (5-36).

Pharmacology

Anti-urease assay (in vitro)

Reaction mixtures comprising 25 μl of jack bean urease enzyme (10 mg/ml) solution and 55 μl of buffers containing 100 mmol urea were incubated with 5 μl of the test compounds (0.5-500 mmol concentration) at 30 °C for 15 min in 96-well plates. Urease activity was determined by measuring ammonia production using the indophenols method as described by Weatherburn [27]. Briefly, 45 μl each of phenol reagent (1% w/v phenol and 0.005% w/v sodium nitroprusside) and 70 μl of alkali reagent (0.5% w/v NaOH and 0.1% active chloride NaOCl) were added to each well. The increasing absorbance at 630 nm was measured after 50 min using a microplate reader (Multimode plate reader, VARIOSKAN, Germany). All the reactions were performed in triplicate in a final volume of 200 μl . The entire assays were performed at pH 6.8 [28]. Percentage inhibition was calculated from the formula:

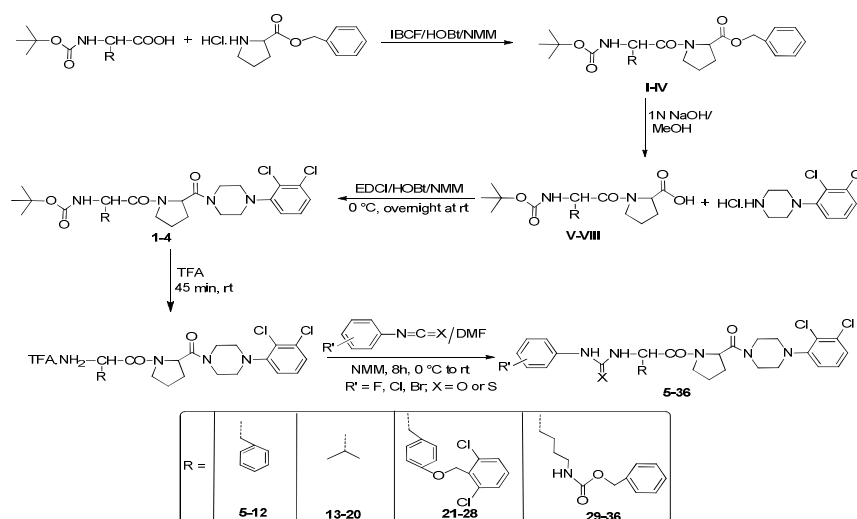
$$\% \text{ Inhibition} = 100 - \frac{\text{OD testwell}}{\text{OD control}} \times 100$$

RESULTS AND DISCUSSION

Chemistry

In the present study, we have synthesized proline containing dipeptides and conjugated to 2,3-dichlorophenyl piperazine. These dipeptide-heterocyclic conjugates were further derivatized into ureas and thioureas by reacting with substituted phenyl isocyanates/isothiocyanates (Scheme).

Yields of the compounds were found to be >85% and were characterized by IR, $^1\text{H NMR}$, mass spectroscopic techniques. IR spectra of ureas and thioureas exhibited peaks at $\nu \sim 1620 \text{ cm}^{-1}$ and $\sim 2040 \text{ cm}^{-1}$ for C=O and C=S respectively. $^1\text{H NMR}$ spectra showed singlet for $-\text{NH}$ at $\delta \sim 8.75$ and multiples for another $-\text{NH}$ proton at $\delta \sim 8.31$ for urea derivatives. On the other hand, δ singlet at ~ 9.70 for $-\text{NH}$ and multiplet at $\delta \sim 8.49$ for NH proton was observed for thiourea derivatives. Further, all other peaks were exactly matching the structure. The physical and mass; IR and NMR data of all the synthesized compounds are presented in table 1 and 2 respectively.



Scheme: Synthesis of ureido and thioureido derivatives of peptide conjugated heterocycle

Table 1: Physical and mass data of synthesized urea/thiourea derivatives (5-36)

Entry	R	X	R _f ^a	R _f ^b	Yield (%)	MP (°C)	Mol. For.	Mass (M ⁺ +1)
5	3F	O	0.53	0.61	91	75	C ₃₁ H ₃₂ Cl ₂ FN ₅ O ₃	613.50
6	4F	O	0.55	0.63	88	73	C ₃₁ H ₃₂ Cl ₂ FN ₅ O ₃	613.54
7	4F	S	0.51	0.59	94	58	C ₃₁ H ₃₂ Cl ₂ FN ₅ O ₂ S	629.55
8	3Cl	O	0.49	0.58	85	72	C ₃₁ H ₃₂ Cl ₃ N ₅ O ₃	629.10
9	3Cl	S	0.45	0.56	84	58	C ₃₁ H ₃₂ Cl ₃ N ₅ O ₂ S	646.10
10	4Cl	S	0.42	0.57	86	60-62	C ₃₁ H ₃₂ Cl ₃ N ₅ O ₂ S	646.12
11	3Br	O	0.52	0.65	88	68	C ₃₁ H ₃₂ BrCl ₂ N ₅ O ₃	674.12
								676.15
12	3Br	S	0.50	0.62	90	55	C ₃₁ H ₃₂ BrCl ₂ N ₅ O ₃	690.02
								692.09
13	3F	O	0.49	0.58	92	58	C ₂₇ H ₃₂ Cl ₂ FN ₅ O ₃	565.45
14	4F	O	0.45	0.56	83	58-60	C ₂₇ H ₃₂ Cl ₂ FN ₅ O ₃	565.52
15	4F	S	0.43	0.53	86	Gum	C ₂₇ H ₃₂ Cl ₂ FN ₅ O ₂ S	581.58
16	3Cl	O	0.47	0.58	81	Gum	C ₂₇ H ₃₂ Cl ₃ N ₅ O ₃	581.90
17	3Cl	S	0.45	0.55	84	Gum	C ₂₇ H ₃₂ Cl ₃ N ₅ O ₂ S	598.02
18	4Cl	S	0.44	0.55	88	Gum	C ₂₇ H ₃₂ Cl ₃ N ₅ O ₂ S	598.05
19	3Br	O	0.46	0.59	90	55	C ₂₇ H ₃₂ BrCl ₂ N ₅ O ₃	625.35
								627.42
20	3Br	S	0.44	0.57	91	Gum	C ₂₇ H ₃₂ BrCl ₂ N ₅ O ₂ S	641.50
								643.38
21	3F	O	0.60	0.71	92	85-87	C ₃₈ H ₃₆ Cl ₄ FN ₅ O ₄	788.50
22	4F	O	0.63	0.70	90	90	C ₃₈ H ₃₆ Cl ₄ FN ₅ O ₄	788.48
23	4F	S	0.58	0.66	95	75-77	C ₃₈ H ₃₆ Cl ₄ FN ₅ O ₃ S	804.63
24	3Cl	O	0.63	0.71	88	65	C ₃₈ H ₃₆ Cl ₅ N ₅ O ₄	804.85
25	3Cl	S	0.59	0.68	89	62-64	C ₃₈ H ₃₆ Cl ₅ N ₅ O ₃ S	821.09
26	4Cl	S	0.57	0.65	86	65	C ₃₈ H ₃₆ Cl ₅ N ₅ O ₃ S	821.12
27	3Br	O	0.61	0.67	88	82	C ₃₈ H ₃₆ BrCl ₄ N ₅ O ₄	849.51
								851.21
28	3Br	S	0.57	0.63	90	70	C ₃₈ H ₃₆ BrCl ₄ N ₅ O ₃ S	865.58
								867.61
29	3F	O	0.49	0.57	80	72	C ₃₆ H ₄₁ Cl ₂ FN ₆ O ₅	728.63
30	4F	O	0.45	0.59	84	75	C ₃₆ H ₄₁ Cl ₂ FN ₆ O ₅	727.71
31	4F	S	0.42	0.55	86	60-62	C ₃₆ H ₄₁ Cl ₂ FN ₆ O ₄ S	744.75
32	3Cl	O	0.44	0.58	81	65	C ₃₆ H ₄₁ Cl ₃ N ₆ O ₅	745.12
33	3Cl	S	0.45	0.55	87	58	C ₃₆ H ₄₁ Cl ₃ N ₆ O ₄ S	761.17
34	4Cl	S	0.47	0.56	84	55-57	C ₃₆ H ₄₁ Cl ₃ N ₆ O ₄ S	761.20
35	3Br	O	0.49	0.58	82	56-58	C ₃₆ H ₄₁ BrCl ₂ N ₆ O ₅	789.58
								791.12
36	3Br	S	0.41	0.53	90	54	C ₃₆ H ₄₁ Cl ₂ N ₆ O ₄ S	805.58
								807.44

R_f= Retention factor, MP= Melting point, ^b= The chemical shift values for the fragment 'Heterocycle' of the compounds 5-36 are almost same as obtained for compound 5

Table 2: IR and NMR values of urea and thiourea derivatives of dipeptides conjugated PZN

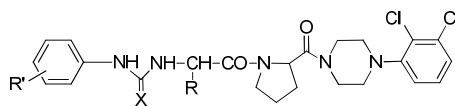
Entry	R/X	IR ν, cm ⁻¹			¹ H NMR (DMSO d ₆ , δ ppm)
		CO	CS	NH	
5	3F(O)	1622		3328	Urea = 8.24 (1H, s, -NH), 6.54-7.42 (4H, m, Ar-H); Phe = 7.93 (1H, s, -NH), 6.54-7.44 (5H, m, Ar-H), 4.59-4.92 (1H, m, - ^α CH), 3.42-3.56 (2H, d, ^β CH ₂); Pro = 3.86-3.94 (1H, t, - ^α CH), 1.68-2.18 (4H, m, - ^{βγ} CH ₂), 3.41-3.52 (2H, t, - ^δ CH ₂) Heterocycle ^b = 3.26-3.37 (4H, m, -CH ₂); 3.41-3.45 (4H, m, -CH ₂); 6.54-7.44 (3H, m, Ar-H).
6	4F(O)	1624		3324	Urea = 8.19 (1H, s, -NH), 6.52-7.41 (4H, m, Ar-H); Phe = 7.91 (1H, s, -NH); 6.66-7.58 (5H, m, Ar-H), 4.65-4.84 (1H, m, - ^α CH), 3.56-3.66 (2H, d, ^β CH ₂); Pro = 3.82-3.91 (1H, t, - ^α CH), 1.64-2.28 (4H, m, - ^{βγ} CH ₂), 3.38-3.50 (2H, t, - ^δ CH ₂)
7	4F(S)	1620	2125	3325	Thiourea = 8.79 (1H, s, -NH), 6.52-7.43 (4H, m, Ar-H); Phe = 7.93 (1H, s, NH), 6.74-7.63 (5H, m, Ar-H), 4.83-4.89 (1H, m, - ^α CH), 3.57-3.70 (2H, d, ^β CH ₂); Pro = 3.79-3.94 (1H, t, - ^α CH), 3.38-3.48 (2H, t, - ^δ CH ₂), 1.68-2.26 (4H, m, - ^{βγ} CH ₂)
8	3Cl(O)	1618		3319	Urea = 8.29 (1H, s, -NH), 6.50-7.38 (4H, m, Ar-H); Phe = 7.96 (1H, s, NH); 6.54-7.48 (5H, m, Ar-H), 4.52-4.84 (1H, m, - ^α CH), 3.41-3.56 (2H, d, ^β CH ₂); Pro = 3.79-3.94 (1H, t, - ^α CH), 1.62-2.28 (4H, m, - ^{βγ} CH ₂), 3.41-3.52 (2H, t, - ^δ CH ₂)
9	3Cl(S)	1625	2132	3328	Thiourea = 8.82 (1H, s, -NH), 6.55-7.48 (4H, m, Ar-H); Phe = 7.95 (1H, s, NH), 6.55-7.48 (5H, m, Ar-H), 4.55-4.83 (1H, m, - ^α CH), 3.41-3.50 (2H, d, ^β CH ₂); Pro = 3.74-3.95 (1H, t, - ^α CH), 3.43-3.52 (2H, t, - ^δ CH ₂), 1.68-2.16 (4H, m, - ^{βγ} CH ₂)
10	4Cl(S)	1624	2127	3320	Thiourea = 8.74 (1H, s, -NH), 6.52-7.43 (4H, m, Ar-H); Phe = 7.89 (1H, s, NH), 6.52-7.43 (5H, m, Ar-H), 4.53-4.91 (1H, m, - ^α CH), 3.41-3.51 (2H, d, ^β CH ₂); Pro = 3.79-3.92 (1H, t, - ^α CH), 1.64-2.12 (4H, m, - ^{βγ} CH ₂), 3.39-3.46 (2H, t, - ^δ CH ₂)
11	3Br(O)	1620		3325	Urea = 8.27 (1H, s, -NH), 6.56-7.48 (4H, m, Ar-H); Phe = 7.92 (1H, s, NH), 6.56-7.48 (5H, m, Ar-H), 4.50-4.94 (1H, m, - ^α CH), 3.46-3.56 (2H, d, ^β CH ₂); Pro = 3.74-3.92 (1H, t, - ^α CH), 3.41-3.50 (2H, t, - ^δ CH ₂), 1.68-2.18 (4H, m, - ^{βγ} CH ₂)
12	3Br(S)	1624	2123	3324	Thiourea = 8.66 (1H, s, -NH), 6.58-7.43 (4H, m, Ar-H); Phe = 7.92 (1H, s, NH), 4.53-4.83 (1H, m, - ^α CH), 3.41-3.50 (2H, d, ^β CH ₂), 6.58-7.43 (5H, m, Ar-H);

13	3F(O)	1621	3318	Pro = 3.79-3.94 (1H, t, $-\alpha$ CH), 1.68-2.26 (4H, m, $-\beta\gamma$ CH ₂), 3.43-3.51 (2H, t, $-\delta$ CH ₂) Urea = 8.79 (1H, s, -NH), 7.11-7.61 (4H, m, Ar-H); Val = 7.70-7.72 (1H, d, NH), 4.33-4.52 (1H, t, $-\alpha$ CH), 1.86-1.89 (1H, m, $-\beta$ CH), 1.03-1.04 (6H, d, (CH ₃) ₂); Pro = 3.73-3.83 (1H, t, $-\alpha$ CH), 2.97-3.52 (2H, t, $-\delta$ CH ₂), 1.91-2.95 (4H, m, $-\beta\gamma$ CH ₂)	
14	4F(O)	1628	2132	3332	Urea = 8.81 (1H, s, -NH), 7.11-7.61 (4H, m, Ar-H); Val = 7.72-7.75 (1H, d, NH), 4.44-4.52 (1H, t, $-\alpha$ CH), 1.86-1.89 (1H, m, $-\beta$ CH), 1.04-1.09 (6H, d, (CH ₃) ₂); Pro = 4.02-4.07 (1H, t, $-\alpha$ CH), 1.06-1.31 (4H, m, $-\beta\gamma$ CH ₂), 3.40 (2H, t, $-\delta$ CH ₂)
15	4F(S)	1618	2124	3328	Thiourea = 8.96 (1H, s, -NH), 6.84-7.38 (4H, m, Ar-H); Val = 7.69-7.70 (1H, d, NH), 4.33-4.49 (1H, t, $-\alpha$ CH), 1.86-1.92 (1H, m, $-\beta$ CH), 1.03-1.07 (6H, d, (CH ₃) ₂); Pro = 3.72-3.80 (1H, t, $-\alpha$ CH), 2.97-3.52 (2H, t, $-\delta$ CH ₂), 1.90-2.91 (4H, m, $-\beta\gamma$ CH ₂)
16	3Cl(O)	1620	3325	Urea = 8.75 (1H, s, -NH), 6.82-7.31 (4H, m, Ar-H); Val = 7.73-7.75 (1H, d, NH), 4.34-4.42 (1H, t, $-\alpha$ CH), 1.86-1.89 (1H, m, $-\beta$ CH), 1.04-1.09 (6H, d, (CH ₃) ₂); Pro = 3.70-3.79 (1H, t, $-\alpha$ CH), 2.95-3.40 (2H, t, $-\delta$ CH ₂), 1.92-2.91 (4H, m, $-\beta\gamma$ CH ₂)	
17	3Cl(S)	1624	2124	3329	Thiourea = 8.98 (1H, s, -NH), 6.84-7.22 (4H, m, Ar-H); Val = 7.70-7.72 (1H, d, NH), 4.33-4.44 (1H, t, $-\alpha$ CH), 1.88-1.90 (1H, m, $-\beta$ CH), 1.03-1.04 (6H, d, (CH ₃) ₂); Pro = 3.73-3.83 (1H, t, $-\alpha$ CH), 2.97-3.52 (2H, t, $-\delta$ CH ₂), 1.91-2.95 (4H, m, $-\beta\gamma$ CH ₂)
18	4Cl(S)	1620	2127	3320	Thiourea = 8.94 (1H, s, -NH), 6.81-7.32 (4H, m, Ar-H); Val = 7.72-7.75 (1H, d, NH), 4.36-4.49 (1H, t, $-\alpha$ CH), 1.85-1.92 (1H, m, $-\beta$ CH), 1.04-1.07 (6H, d, (CH ₃) ₂); Pro = 3.72-3.81 (1H, t, $-\alpha$ CH), 2.98-3.52 (2H, t, $-\delta$ CH ₂), 1.93-2.89 (4H, m, $-\beta\gamma$ CH ₂)
19	3Br(O)	1630	3317	Urea = 8.78 (1H, s, -NH), 6.81-7.31 (4H, m, Ar-H); Val = 7.71-7.73 (1H, d, NH), 4.34-4.42 (1H, t, $-\alpha$ CH), 1.85-1.88 (1H, m, $-\beta$ CH), 1.04-1.08 (6H, d, (CH ₃) ₂); Pro = 3.75-3.85 (1H, t, $-\alpha$ CH), 2.89-3.40 (2H, t, $-\delta$ CH ₂), 1.96-2.93 (4H, m, $-\beta\gamma$ CH ₂)	
20	3Br(S)	1628	2122	3320	Thiourea = 8.94 (1H, s, -NH), 6.83-7.32 (4H, m, Ar-H); Val = 7.69-7.71 (1H, d, NH), 4.36-4.49 (1H, t, $-\alpha$ CH), 1.86-1.95 (1H, m, $-\beta$ CH), 1.04-1.06 (6H, d, (CH ₃) ₂); Pro = 3.71-3.79 (1H, t, $-\alpha$ CH), 2.96-3.52 (2H, t, $-\delta$ CH ₂), 1.88-2.89 (4H, m, $-\beta\gamma$ CH ₂)
21	3F(O)	1625	3331	Urea = 8.97 (1H, s, -NH), 6.70-7.90 (4H, m, Ar-H); Tyr = 8.65 (1H, s, NH), 6.70-7.90 (4H, m, Ar-H), 4.82-4.92 (1H, m, $-\alpha$ CH), 3.67-3.76 (2H, d, $-\beta$ CH ₂); 2,6-Cl ₂ -Bzl = 6.70-7.90 (3H, m, Ar-H), 5.19 (2H, s, CH ₂); Pro = 3.36-3.49 (1H, t, $-\alpha$ CH), 2.90-3.16 (2H, t, $-\delta$ CH ₂), 1.67-1.89 (4H, m, $-\beta\gamma$ CH ₂)	
22	4F(O)	1624	3328	Urea = 8.95 (1H, s, -NH), 6.69-7.88 (4H, m, Ar-H); Tyr = 8.60 (1H, s, NH), 6.69-7.88 (4H, m, Ar-H), 4.82-4.92 (1H, m, $-\alpha$ CH), 3.65-3.72 (2H, d, $-\beta$ CH ₂); 2,6-Cl ₂ -Bzl = 6.69-7.88 (3H, m, Ar-H); 5.14 (2H, s, CH ₂); Pro = 3.35-3.46 (1H, t, $-\alpha$ CH), 2.90-3.16 (2H, t, $-\delta$ CH ₂), 1.68-1.88 (4H, m, $-\beta\gamma$ CH ₂)	
23	4F(S)	1623	2125	3322	Thiourea = 9.08 (1H, s, -NH), 6.74-7.89 (4H, m, Ar-H); Tyr = 8.62 (1H, s, NH), 6.74-7.89 (4H, m, Ar-H), 4.84-4.95 (1H, m, $-\alpha$ CH), 3.64-3.78 (2H, d, $-\beta$ CH ₂); 2,6-Cl ₂ -Bzl = 6.74-7.89 (3H, m, Ar-H), 5.14 (2H, s, CH ₂); Pro = 3.30-3.45 (1H, t, $-\alpha$ CH), 2.92-3.14 (2H, t, $-\delta$ CH ₂), 1.60-1.82 (4H, m, $-\beta\gamma$ CH ₂)
24	3Cl(O)	1620	3329	Urea = 8.95 (1H, s, -NH), 6.75-7.91 (4H, m, Ar-H); Tyr = 8.62 (1H, s, NH), 6.75-7.91 (4H, m, Ar-H), 4.82-4.92 (1H, m, $-\alpha$ CH), 3.68-3.75 (2H, d, $-\beta$ CH ₂); 2,6-Cl ₂ -Bzl = 6.75-7.91 (3H, m, Ar-H), 5.19 (2H, s, CH ₂); Pro = 3.33-3.45 (1H, t, $-\alpha$ CH), 2.92-3.16 (2H, t, $-\delta$ CH ₂), 1.64-1.86 (4H, m, $-\beta\gamma$ CH ₂)	
25	3Cl(S)	1619	2125	3330	Thiourea = 9.13 (1H, d, -NH), 6.74-7.90 (4H, m, Ar-H); Tyr = 8.62 (1H, s, NH), 6.74-7.90 (4H, m, Ar-H), 4.80-4.92 (1H, m, $-\alpha$ CH), 3.64-3.74 (2H, d, $-\beta$ CH ₂); 2,6-Cl ₂ -Bzl = 6.74-7.90 (3H, m, Ar-H), 5.15 (2H, s, CH ₂); Pro = 3.32-3.46 (1H, t, $-\alpha$ CH), 2.92-3.18 (2H, t, $-\delta$ CH ₂), 1.62-1.85 (4H, m, $-\beta\gamma$ CH ₂)
26	4Cl(S)	1622	2126	3328	Thiourea = 9.18 (1H, s, -NH), 6.74-7.90 (4H, m, Ar-H); Tyr = 8.68 (1H, s, NH), 6.74-7.90 (4H, m, Ar-H), 4.81-4.90 (1H, m, $-\alpha$ CH), 3.66-3.74 (2H, d, $-\beta$ CH ₂); 2,6-Cl ₂ -Bzl = 6.74-7.90 (3H, m, Ar-H), 5.14 (2H, s, CH ₂); Pro = 3.33-3.45 (1H, t, $-\alpha$ CH), 2.92-3.14 (2H, t, $-\delta$ CH ₂), 1.67-1.82 (4H, m, $-\beta\gamma$ CH ₂)
27	3Br(O)	1624	3321	Urea = 8.97 (1H, s, -NH), 6.70-7.90 (4H, m, Ar-H); Tyr = 8.65 (1H, s, NH), 6.70-7.90 (4H, m, Ar-H), 4.81-4.90 (1H, m, $-\alpha$ CH), 3.66-3.76 (2H, d, $-\beta$ CH ₂); 2,6-Cl ₂ -Bzl = 6.70-7.90 (3H, m, Ar-H); 5.17 (2H, s, CH ₂); Pro = 3.35-3.48 (1H, t, $-\alpha$ CH), 2.94-3.16 (2H, t, $-\delta$ CH ₂), 1.68-1.88 (4H, m, $-\beta\gamma$ CH ₂)	
28	3Br(S)	1625	2130	3325	Thiourea = 9.15 (1H, s, -NH), 6.71-7.89 (4H, m, Ar-H); Tyr = 8.63 (1H, s, NH), 6.71-7.89 (4H, m, Ar-H), 4.82-4.95 (1H, m, $-\alpha$ CH), 3.64-3.74 (2H, d, $-\beta$ CH ₂); 2,6-Cl ₂ -Bzl = 6.71-7.89 (3H, m, Ar-H), 5.15 (2H, s, CH ₂); Pro = 3.34-3.47 (1H, t, $-\alpha$ CH), 2.92-3.14 (2H, t, $-\delta$ CH ₂), 1.60-1.82 (4H, m, $-\beta\gamma$ CH ₂)
29	3F(O)	1617	3328	Urea = 8.34 (1H, s, -NH), 7.11-7.66 (4H, m, Ar-H); Lys = 8.08-8.12 (1H, m, -NH), 4.12-4.15 (1H, m, $-\alpha$ CH), 3.17-3.21 (2H, m, $-\epsilon$ CH ₂), 1.70-2.24 (2H, m, $-\beta$ CH ₂), 1.58-1.62 (2H, m, $-\delta$ CH ₂), 1.40-1.48 (2H, m, $-\gamma$ CH ₂); Z = 7.81-7.92 (1H, t, NH), 7.11-7.66 (5H, m, Ar-H), 5.19 (2H, s, CH ₂); Pro = 3.33-3.46 (1H, t, $-\alpha$ CH), 2.95-3.16 (2H, t, $-\delta$ CH ₂), 1.62-1.96 (4H, m, $-\beta\gamma$ CH ₂)	
30	4F(O)	1628	3331	Urea = 8.38 (1H, s, -NH), 7.12-7.68 (4H, m, Ar-H); Lys = 8.10-8.14 (1H, m, NH), 4.17-4.21 (1H, m, $-\alpha$ CH), 3.18-3.21 (2H, m, $-\epsilon$ CH ₂), 1.72-2.24 (2H, m, $-\beta$ CH ₂), 1.55-1.63 (2H, m, $-\delta$ CH ₂), 1.43-1.47 (2H, m, $-\gamma$ CH ₂); Z = 7.71-7.74 (1H, t, NH), 7.12-7.68 (5H, m, Ar-H), 5.19 (2H, s, CH ₂); Pro = 3.31-3.45 (1H, t, $-\alpha$ CH), 1.62-1.96 (4H, m, $-\beta\gamma$ CH ₂), 2.98-3.26 (2H, t, $-\delta$ CH ₂)	
31	4F(S)	1624	2130	3327	Thiourea = 9.85 (1H, s, -NH), 7.11-7.68 (4H, m, Ar-H); Lys = 8.29-8.32 (1H, m, -NH), 4.13-4.16 (1H, m, $-\alpha$ CH), 3.17-3.24 (2H, m, $-\epsilon$ CH ₂), 1.77-2.14 (2H, m, $-\beta$ CH ₂), 1.53-1.61 (2H, m, $-\delta$ CH ₂), 1.42-1.47 (2H, m, $-\gamma$ CH ₂); Z = 7.80-7.85 (1H, m, NH), 7.11-7.68 (5H, m, Ar-H), 5.16 (2H, s, CH ₂); Pro = 3.34-3.47 (1H, t, $-\alpha$ CH), 1.63-2.16 (4H, m, $-\beta\gamma$ CH ₂), 2.94-3.12 (2H, t, $-\delta$ CH ₂)
32	3Cl(O)	1626	3328	Urea = 8.34 (1H, s, -NH), 7.12-7.69 (4H, m, Ar-H); Lys = 8.10-8.14 (1H, m, NH), 4.17-4.21 (1H, m, $-\alpha$ CH), 3.28-3.41 (2H, m, $-\epsilon$ CH ₂), 1.70-2.24 (2H, m, $-\beta$ CH ₂), 1.65-1.73 (2H, m, $-\delta$ CH ₂), 1.41-1.42 (2H, m, $-\gamma$ CH ₂);	

33	3Cl(S)	1625	2128	3335	Z = 7.82-7.94 (1H, t, NH), 7.12-7.69 (5H, m, Ar-H), 5.17 (2H, s, CH ₂); Pro = 3.35-3.49 (1H, t, - ^α CH), 2.95-3.16 (2H, t, - ^δ CH ₂), 1.65-1.98 (4H, m, - ^{βγ} CH ₂) Thiourea = 9.82 (1H, s, -NH), 7.01-7.68 (4H, m, Ar-H); Lys = 8.29-8.34 (1H, m, NH), 4.13-4.16 (1H, m, - ^α CH), 3.19-3.24 (2H, m, - ^ε CH ₂), 1.72-2.14 (2H, m, - ^β CH ₂), 1.53-1.61 (2H, m, - ^δ CH ₂), 1.39-1.49 (2H, m, - ^γ CH ₂);
34	4Cl(S)	1628	2130	3322	Z = 7.80-7.85 (1H, t, NH), 5.16 (2H, s, CH ₂), 7.01-7.68 (5H, m, Ar-H); Pro = 3.33-3.47 (1H, t, - ^α CH), 2.94-3.22 (2H, t, - ^δ CH ₂), 1.64-1.96 (4H, m, - ^{βγ} CH ₂) Thiourea = 9.80 (1H, s, -NH), 7.08-7.72 (4H, m, Ar-H); Lys = 8.29-8.35 (1H, m, NH), 4.15-4.18 (1H, m, - ^α CH), 3.17-3.24 (2H, m, - ^ε CH ₂), 1.77-2.14 (2H, m, - ^β CH ₂), 1.58-1.61 (2H, m, - ^δ CH ₂), 1.49-1.54 (2H, m, - ^γ CH ₂);
35	3Br(O)	1619		3326	Z = 7.83-7.85 (1H, t, NH), 7.08-7.72 (5H, m, Ar-H), 5.19 (2H, s, CH ₂); Pro = 3.34-3.49 (1H, t, - ^α CH), 2.94-3.12 (2H, t, - ^δ CH ₂), 1.63-1.96 (4H, m, - ^{βγ} CH ₂) Urea = 8.34 (1H, s, -NH), 7.02-7.66 (4H, m, Ar-H); Lys = 8.06-8.08 (1H, m, NH), 4.17-4.21 (1H, m, - ^α CH), 3.18-3.21 (2H, m, - ^ε CH ₂), 1.70-2.24 (2H, m, - ^β CH ₂), 1.59-1.73 (2H, m, - ^δ CH ₂), 1.41-1.44 (2H, m, - ^γ CH ₂);
36	3Br(S)	1624	2130	3321	Z = 7.71-7.64 (1H, t, NH), 7.02-7.66 (5H, m, Ar-H), 5.19 (2H, s, CH ₂); Pro = 3.35-3.49 (1H, t, - ^α CH), 2.98-3.21 (2H, t, - ^δ CH ₂), 1.62-2.06 (4H, m, - ^{βγ} CH ₂) Thiourea = 9.85 (1H, s, -NH), 7.01-7.68 (4H, m, Ar-H); Lys = 8.29-8.32 (1H, m, NH), 4.13-4.16 (1H, m, - ^α CH), 3.19-3.24 (2H, m, - ^ε CH ₂), 1.77-2.14 (2H, m, - ^β CH ₂), 1.73-2.22 (2H, m, - ^δ CH ₂), 1.42-1.50 (2H, m, - ^γ CH ₂); Z = 7.83-7.86 (1H, t, NH), 7.01-7.68 (5H, m, Ar-H), 5.17 (2H, s, CH ₂); Pro = 3.32-3.48 (1H, t, - ^α CH), 2.94-3.12 (2H, t, - ^δ CH ₂), 1.68-2.06 (4H, m, - ^{βγ} CH ₂)

IR = Infrared spectroscopic values, NMR = Nuclear Magnetic Resonance spectroscopic values, DMSO = Dimethyl sulfoxide.

Table 3: Antiurease studies of the synthesized compounds (1-36). Each value represents mean±SD (n=3)



R' = F, Cl, Br; R = Side chain of amino acids; X = O/S

Entry	R	X	Antiurease activity (μM) ^a
1	Boc-FP-PZN		78.5±0.69
2	Boc-VP-PZN		92.1±0.55
3	Boc-Y(2,6CL2Bzl)P-PZN		56.7±0.55
4	Boc-K(Z)P-PZN		32.5±0.92
5	3F	O	23.5±0.18
6	4F	O	15.8±0.13
7	4F	S	6.0±0.11
8	3Cl	O	20.7±0.25
9	3Cl	S	18.2±0.31
10	4Cl	S	15.0±0.15
11	3Br	O	28.0±0.12
12	3Br	S	25.7±0.47
13	3F	O	28.0±0.59
14	4F	O	21.6±0.24
15	4F	S	7.4±0.18
16	3Cl	O	31.8±0.64
17	3Cl	S	30.6±0.96
18	4Cl	S	21.1±0.21
19	3Br	O	45.6±0.95
20	3Br	S	32.5±0.33
21	3F	O	20.0±0.25
22	4F	O	12.0±0.36
23	4F	S	2.0±0.12
24	3Cl	O	7.0±0.21
25	3Cl	S	4.4±0.18
26	4Cl	S	3.0±0.14
27	3Br	O	18.3±0.25
28	3Br	S	16.0±0.26
29	3F	O	27.2±0.76
30	4F	O	18.4±0.41
31	4F	S	3.12±0.08
32	3Cl	O	21.8±0.72
33	3Cl	S	19.2±0.32
34	4Cl	S	8.91±0.17
35	3Br	O	28.4±0.41
36	3Br	S	21.0±0.22
Thiourea			21.0±0.11

^aValues are mean of three determinations, the range of which are <5% of mean in all cases

Pharmacology

The focus of our approach in this work was to evaluate the urease inhibitory activity of the title compounds by *in vitro* method. Data obtained is presented in table 3. Thiourea was used as the reference standard in the assay. Analysis of the data from this study suggests that (1) Most of the synthesized urea and thiourea derivatives with substitution on the phenyl ring were found to be more active than standard thiourea, (2) the activity of the synthesized compounds is dependent upon the nature of amino acids which is bridging the urea/thiourea group and the heterocycle and also position of the substitution on the aromatic ring of urea/thioureas.

Among the four series of dipeptides, Tyr containing dipeptide conjugated heterocyclic derivatives (23-26) were found to be most active followed by Lys dipeptide conjugated heterocyclic compounds (29-36). On the other hand, Phe dipeptide conjugated heterocyclic compounds (5-12) and Val dipeptide conjugated heterocyclic compounds (13-20) have shown moderate activity. The observed results revealed that substitution pattern on the phenyl ring of ureas/thioureas plays very important role on the activity. Compounds which have thiourea group 7, 10, 15, 23, 25, 26 and 31 have exhibited high potent activity with an IC_{50} values of 10.0 ± 0.11 , 15.0 ± 0.15 , 17.4 ± 0.18 , 4.0 ± 0.12 , 7.0 ± 0.21 , 4.4 ± 0.18 , 2.0 ± 0.14 and 3.12 ± 0.08 μM respectively. The remarkable activity of compounds 23, 25, 26 and 31 may be due to electron withdrawing substituent which is more desirable. Further, it was noticed that changing in substitution position on the aryl ring changes the activity. When substituents present at *para* and *meta* position exhibited maximum activity compared to *ortho*. Compounds containing thiourea have emerged as active moieties which indicated that involving 'S' in the design would be more beneficial which could attribute to the electronegative behaviour of 'S' and its ability to react with target group there by increasing the efficiency of the compounds.

The urease inhibition capacity of newly synthesized molecules was compared with the literature reported other studies. Molecules like morpholine [4], metronidazole [6], substituted piperidines [7], Barbiturates [8] and benzoquinone derivatives [9] having IC_{50} values 2.37 μM , 12 μM , 13 μM , 19.38 μM and 27.30 μM respectively. Most of our analogues were found to be more active than the results with those of other studies. Importantly the analogue 23 was nearly 10 fold (2 μM) more potent than the reference standard, 21.0 ± 0.11 μM .

We were interested to give a preliminary mechanistic approach for the ability of the synthesized compounds to inhibit the urease activity i.e., the inhibition of the production of ammonia. Urease is a dinickel containing enzyme surrounded by molecules. In literature, it has been described that urea binds to nickel using the carbonyl oxygen and displaces the water molecules. This further facilitates the urea coordination to the second Ni²⁺ by hydrogen binding. Nucleophilic attack of oxygen (of OH) to carbonyl carbon followed by proton abstraction leads to the elimination of ammonia molecule [29]. More the concentration of ammonia, lesser will be the activity. Thus in the present work substituted urea and thiourea derivatives would not result in the production of ammonia and hence it may be concluded that compounds of the type described in this work may act as more active analogues.

CONCLUSION

In conclusion, four series of dipeptides [FP, VP, Y(2,6-Cl₂-Bzl)P and K(Z)P] conjugated 2,3-dichlorophenyl piperazine were synthesized and converted into urea and thiourea derivatives by reacting with various substituted phenyl isocyanates/isothiocyanates. All the synthesized compounds were evaluated for their urease inhibitory potential. These inhibitors demonstrated significant *in vitro* efficacy as antifreeze agents. Compounds with 'S' and preferably F and Cl substituents at *meta* and *para* positions showed predominant urease inhibitory activity. These results suggest the suitability of the designed molecular frame work for use in the development of anti-urease therapy.

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AUTHORS CONTRIBUTION

D. M. Suyoga Vardhan and D. Channe Gowda involved in the synthesis of urea and thiourea derivatives of dipeptides conjugated 2,3-dichlorophenyl piperazine and their characterization. H. K. Kumara and H. Pavan Kumar contributed in urease inhibition assay.

CONFLICT OF INTERESTS

The authors declare that they have no conflict of interest

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